

# From donor biology to donor health protection: Three (very) short stories

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COLUMBIA UNIVERSITY

*College of Physicians  
and Surgeons*

 **NewYork-Presbyterian**  
The University Hospitals of Columbia and Cornell

# Potential Conflicts of Interest

**Hemanext:**

**Advisory Board**

**Tioma, Inc:**

**Consultant**

# This is the focus of our interest



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**Are there characteristics of the donor that will “make better products” and affect donor health?**

**What do we know about  
refrigerator-stored RBCs?**

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**As storage time increases (FDA criteria):**

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Increasing hemolysis *ex vivo* (<1.0%)

**Infuse free hemoglobin, etc.**

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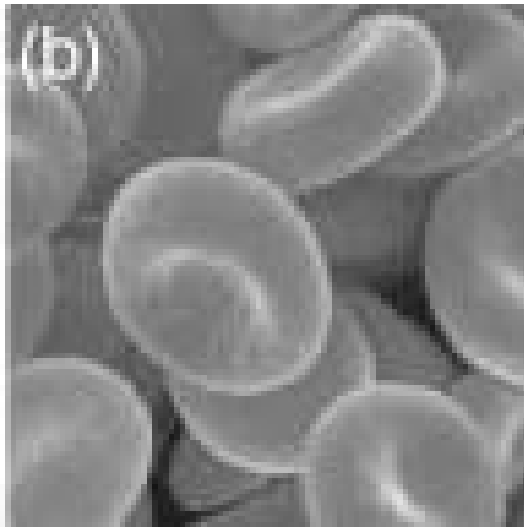
Decreasing 24-hr post-transfusion recovery *in vivo* ( $\sim \geq 75\%$ )

**Less than optimal dose**



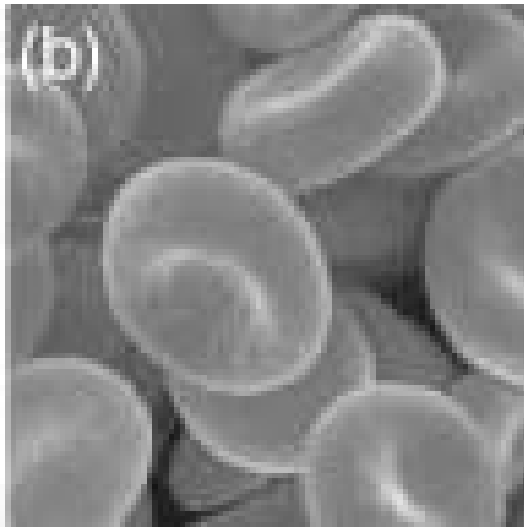
**What happens to the RBCs  
during refrigerated storage?**

# The “RBC storage lesion”



**Day 0**

# The “RBC storage lesion”



**Day 0**



**Day 42**

# The “RBC storage lesion”

- ↓ 2,3-DPG, GSH, ATP
- ↓ Nitric oxide
- ↑ Protein oxidation
- ↑ Membrane- & cytoskeletal-associated hemoglobin
- ↑ Membrane lipid peroxidation
- ↑ Lysophosphatidylcholine species
- ↑ Vesiculation and membrane loss
- ↓ Deformability
- ↑ Phosphatidylserine exposure
- ↓ CD47

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# The “RBC storage lesion”

## Final common pathway?

Metabolic dysfunction & oxidative stress →

↓ Deformability

↑ “Eat me” signals (**phosphatidylserine**)

↓ “Don’t eat me” signals (**CD47**)

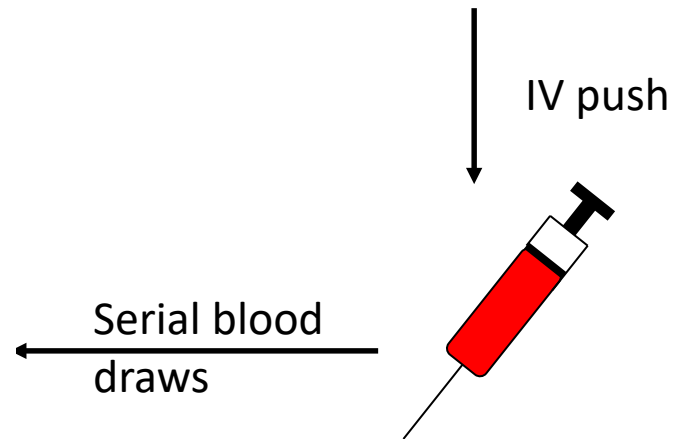
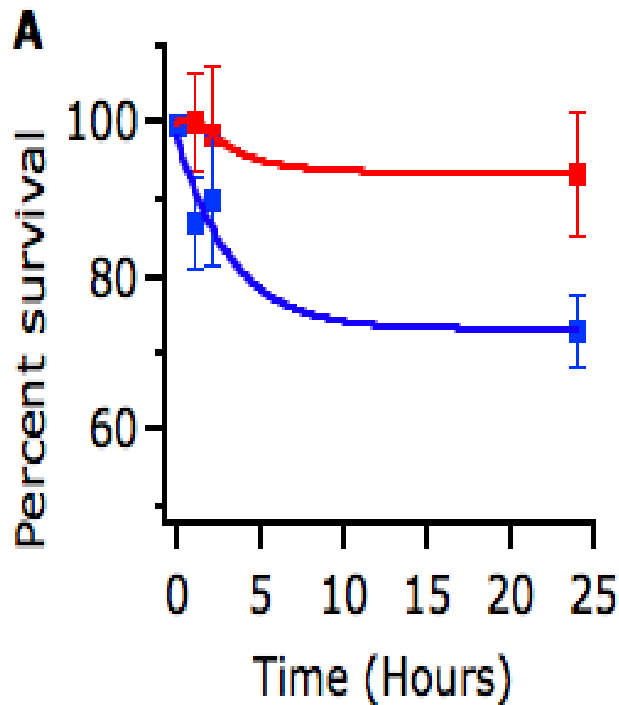
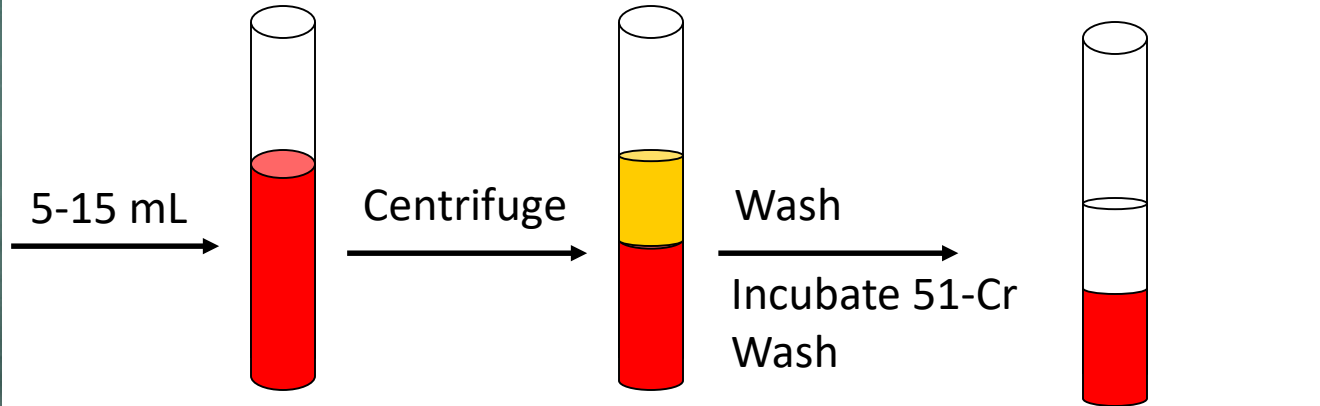
↑ Hemolysis *in vitro*

↑ RBC clearance *in vivo*

Intravascular and extravascular hemolysis

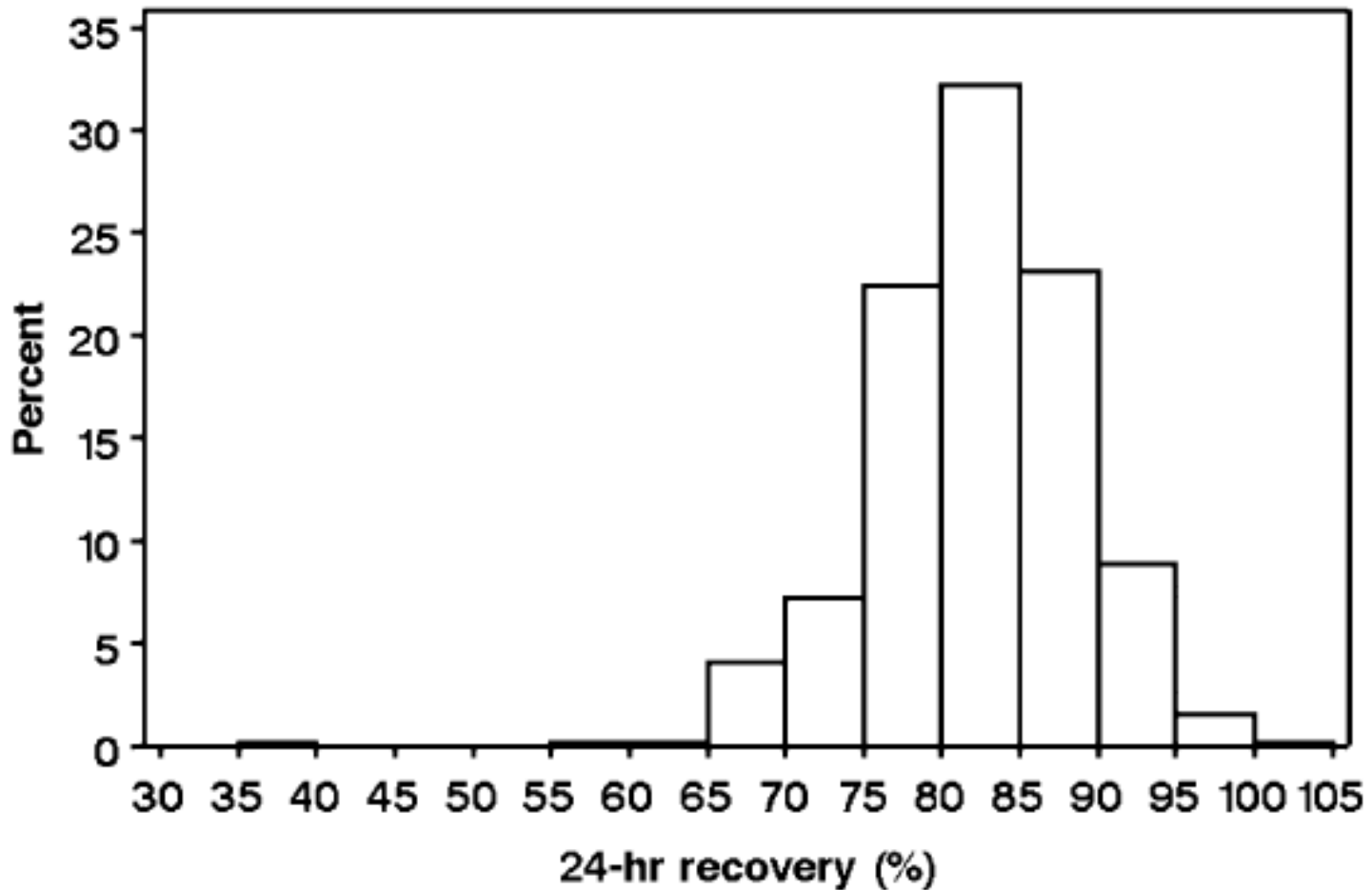
# **RBC Clearance Variability**

# RBC Recovery Study



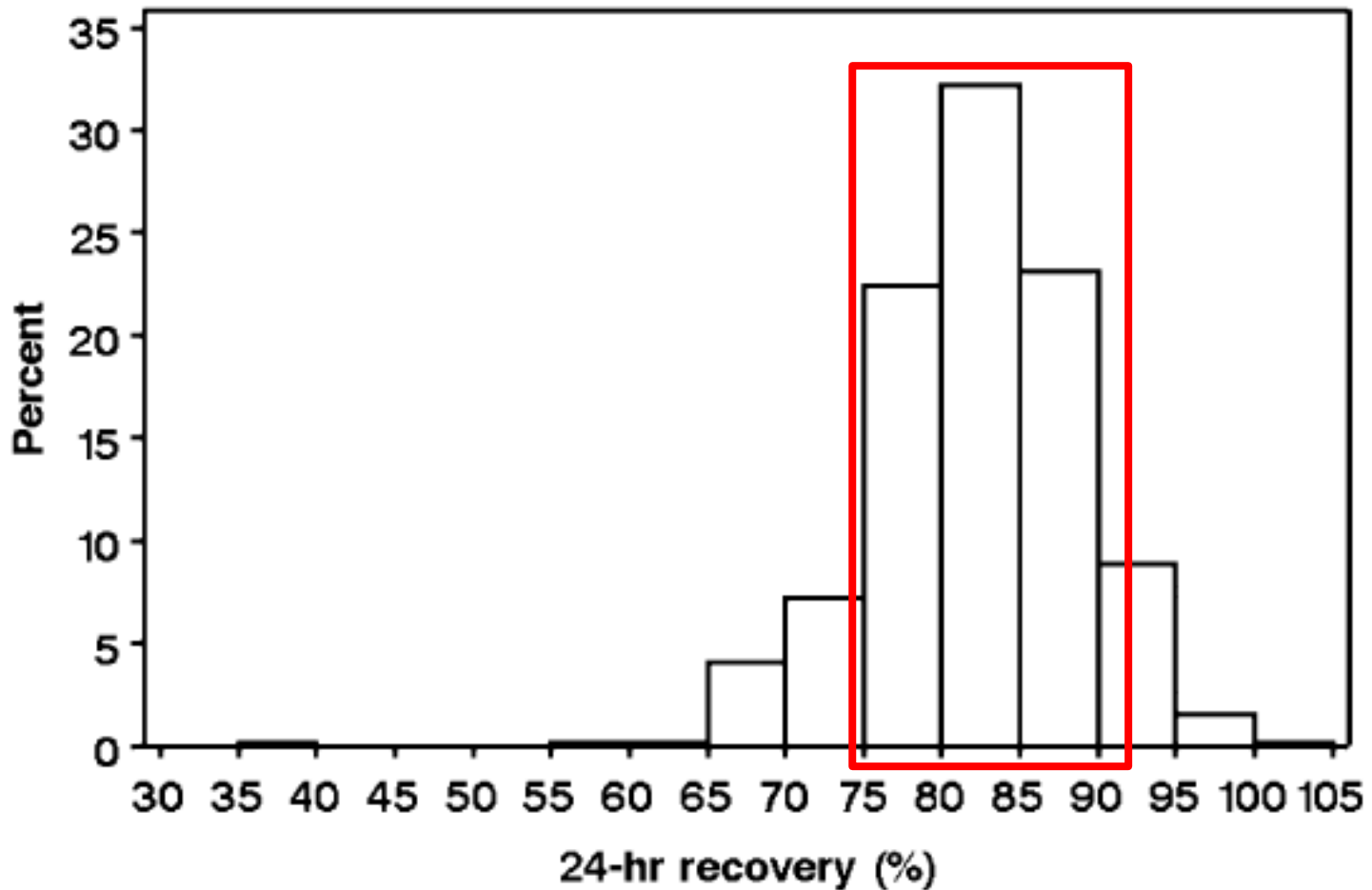


# 24-hr RBC recovery in 641 healthy volunteers



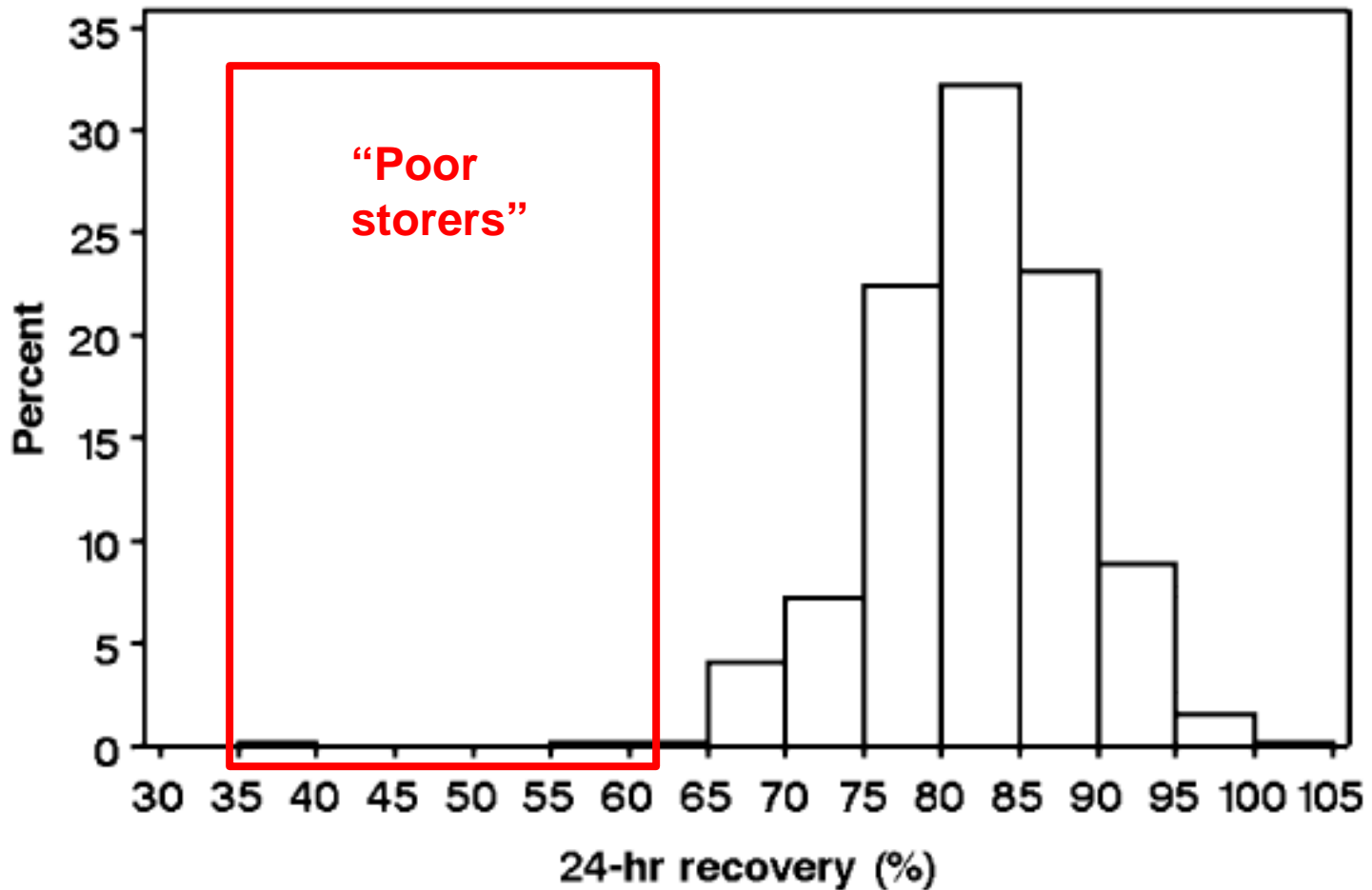
Dumont et al. Transfusion 48:1053-60, 2008.

# 24-hr RBC recovery in 641 healthy volunteers



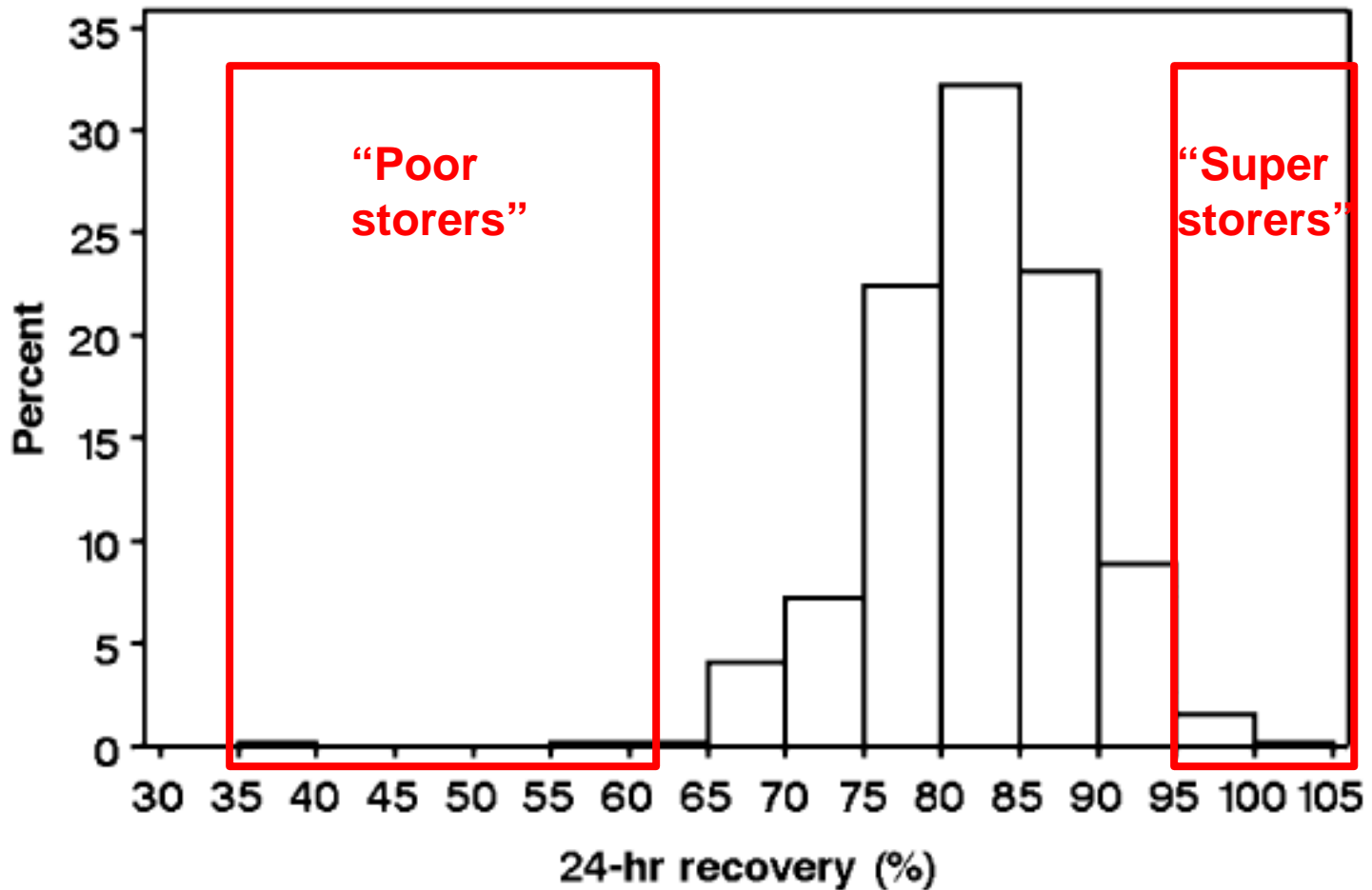
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# 24-hr RBC recovery in 641 healthy volunteers



Dumont et al. Transfusion 48:1053-60, 2008.

**Is “old” blood bad?**

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**Infection?**

**Inflammation?**

**Thrombosis?**

**Mortality?**

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**Not going to talk about these now**

# **Is “old” blood bad?**

**Infection?**

**Inflammation?**

**Thrombosis?**

**Mortality?**

**Not going to talk about these now**

**We can discuss these later, if you  
would like**



# Is “old” blood bad?

↑ RBC refrigerated storage time



↑ RBC storage lesion *in vitro*



↓ RBC recovery *in vivo*

# **Is “old” blood bad?**

**Why is transfusion of less than a full “dose” OK?**

# Is “old” blood bad?

Why is transfusion of less than a full “dose” OK?

Transfused RBCs that don't circulate cannot deliver O<sub>2</sub>

**When do RBCs “go bad”?**

# When do RBCs “go bad”?

The Journal of Clinical Investigation

CLINICAL MEDICINE

## Prolonged red cell storage before transfusion increases extravascular hemolysis

Francesca Rapido,<sup>1,2</sup> Gary M. Brittenham,<sup>3,4</sup> Sheila Bandyopadhyay,<sup>1</sup> Francesca La Carpia,<sup>1</sup> Camilla L'Acqua,<sup>1</sup> Donald J. McMahon,<sup>4</sup> Abdelhadi Rebbaa,<sup>1</sup> Boguslaw S. Wojczyk,<sup>1</sup> Jane Netterwald,<sup>1</sup> Hangli Wang,<sup>1</sup> Joseph Schwartz,<sup>1</sup> Andrew Eisenberger,<sup>4</sup> Mark Soffing,<sup>5</sup> Randy Yeh,<sup>5</sup> Chaitanya Divgi,<sup>5</sup> Yelena Z. Ginzburg,<sup>6</sup> Beth H. Shaz,<sup>6</sup> Sujit Sheth,<sup>7</sup> Richard O. Francis,<sup>1</sup> Steven L. Spitalnik,<sup>1</sup> and Eldad A. Hod<sup>1</sup>

**Journal of Clinical Investigation 127:375-382, 2017**

**60 healthy volunteers enrolled**

**52 completed study**

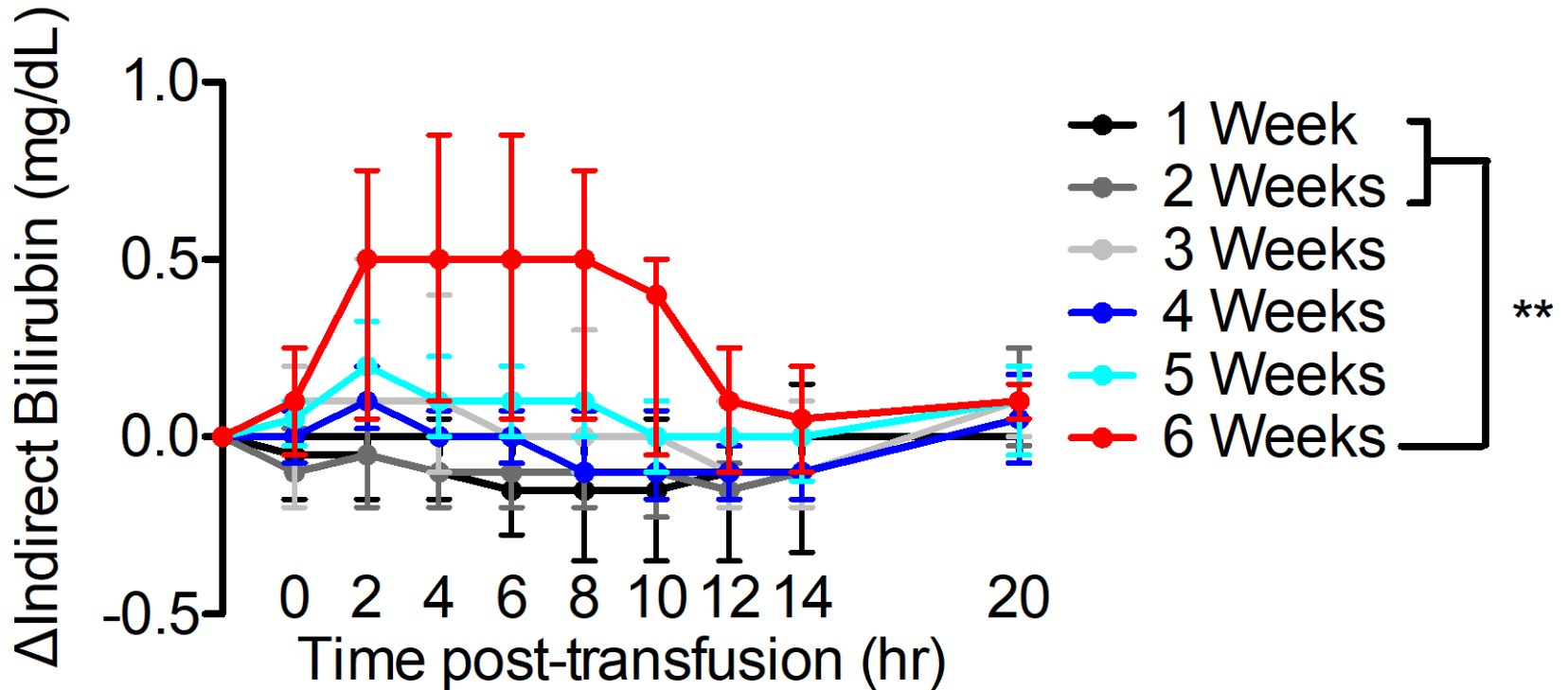
**Randomized to 1, 2, 3, 4, 5, 6 weeks of storage**

**Leukoreduced; AS-3**

**Transfused entire unit**

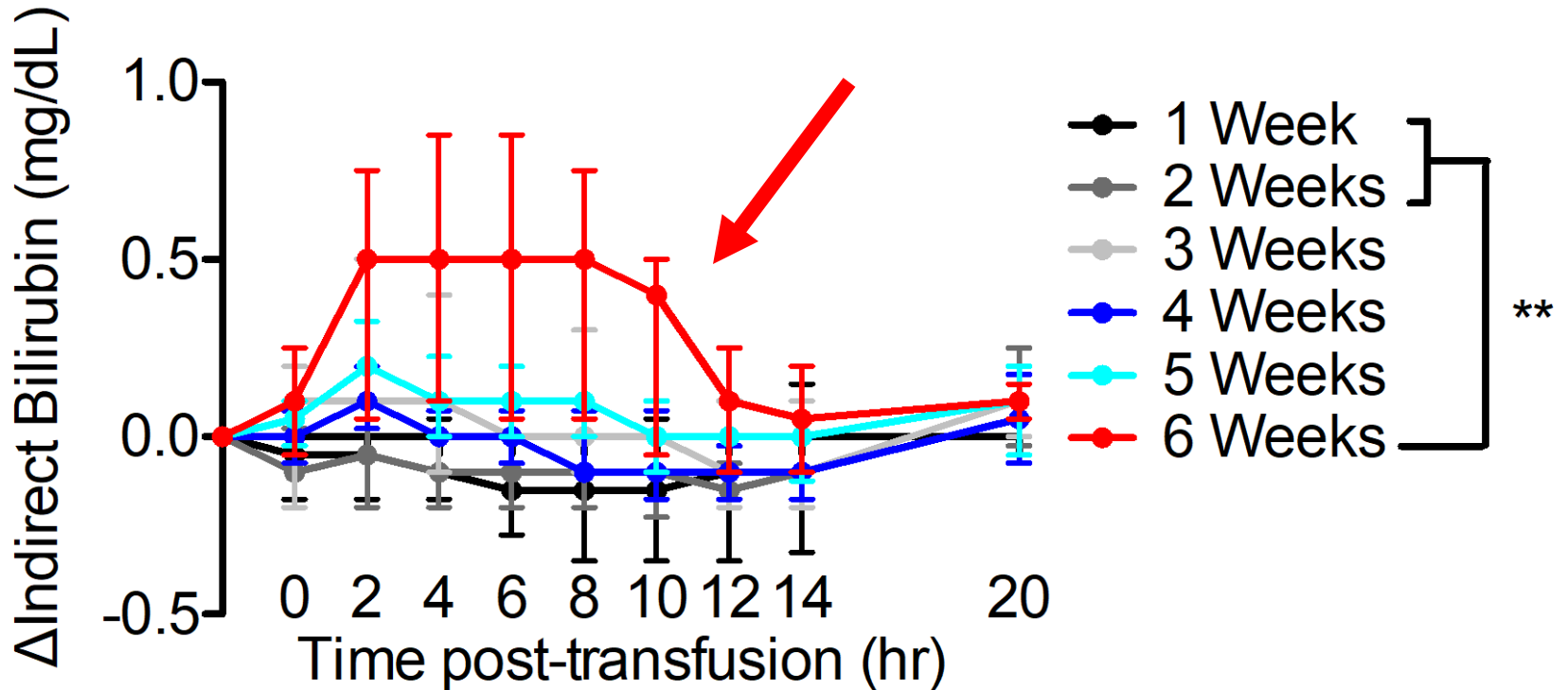
**$^{51}\text{Cr}$ -labeled post-transfusion recovery**

# When do RBCs “go bad”?



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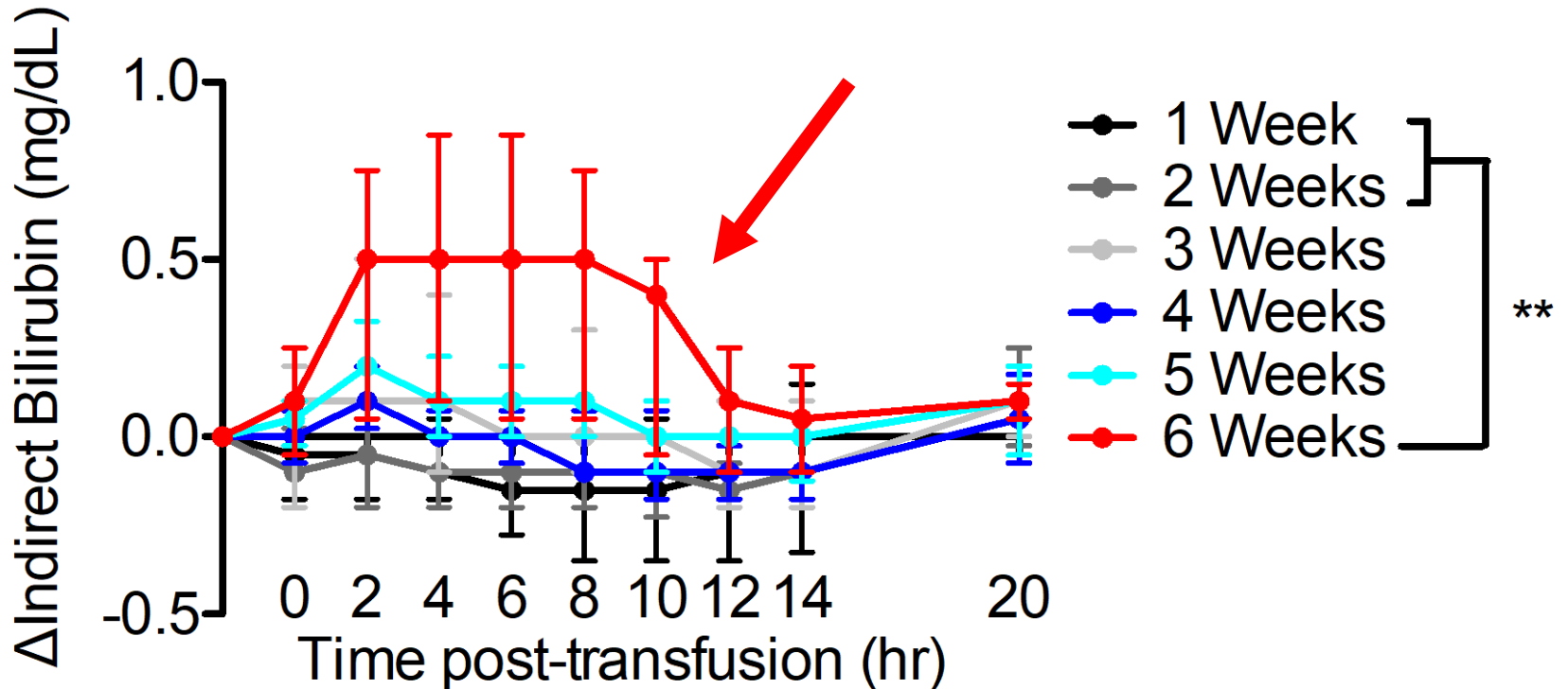
6 Weeks





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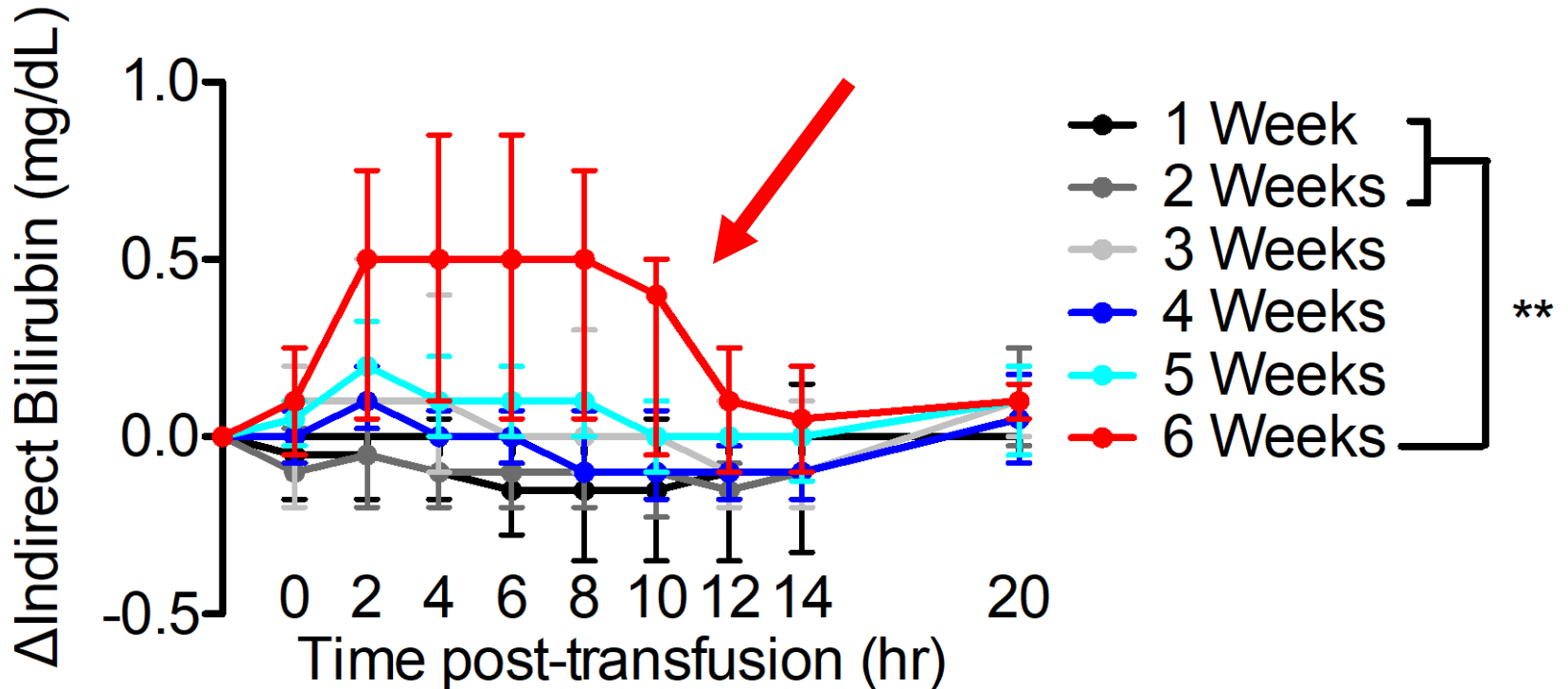
**6 Weeks**



**Similar pattern with serum iron, NTBI, etc.**

# When do RBCs “go bad”?

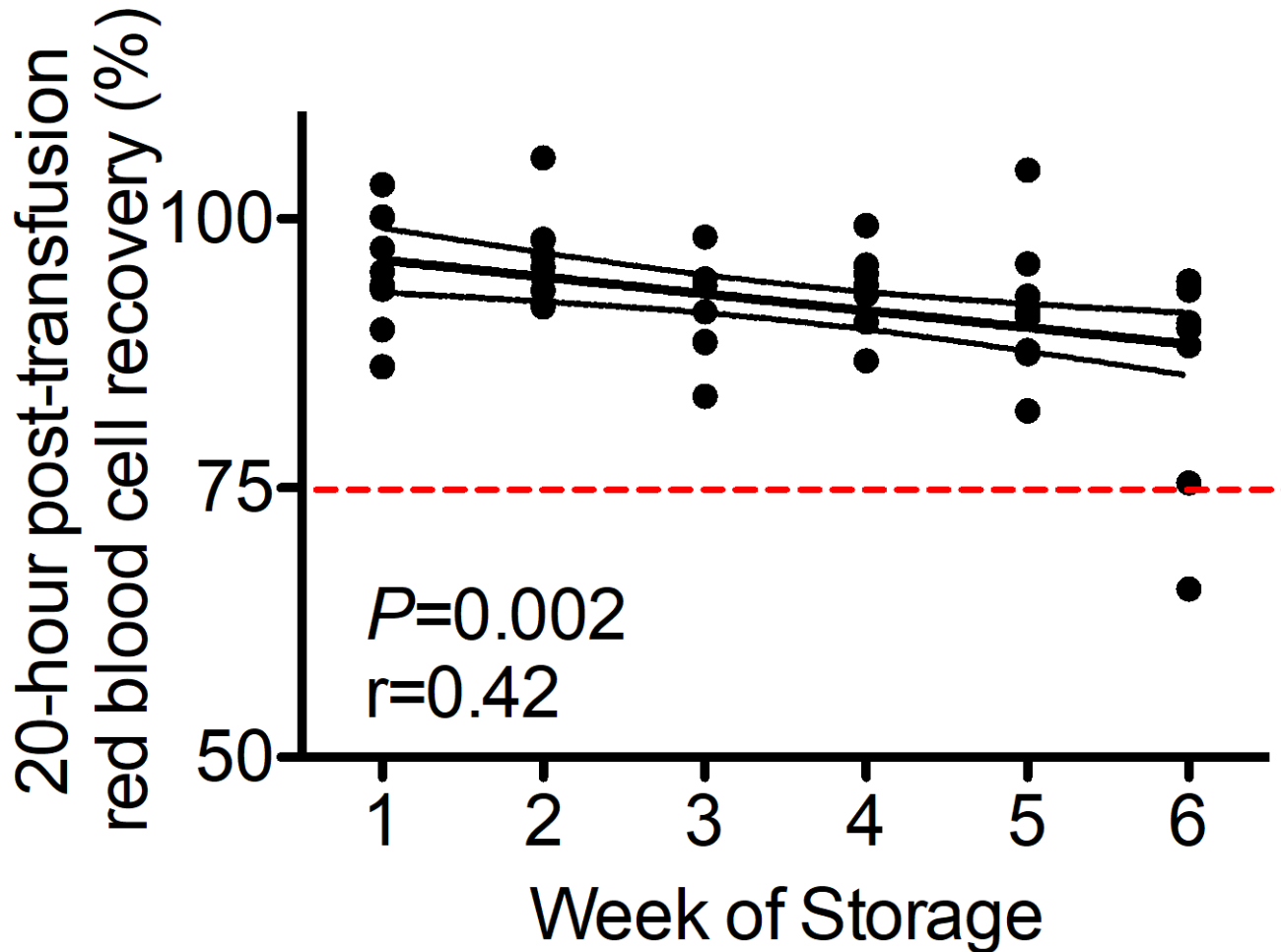
**6 Weeks**



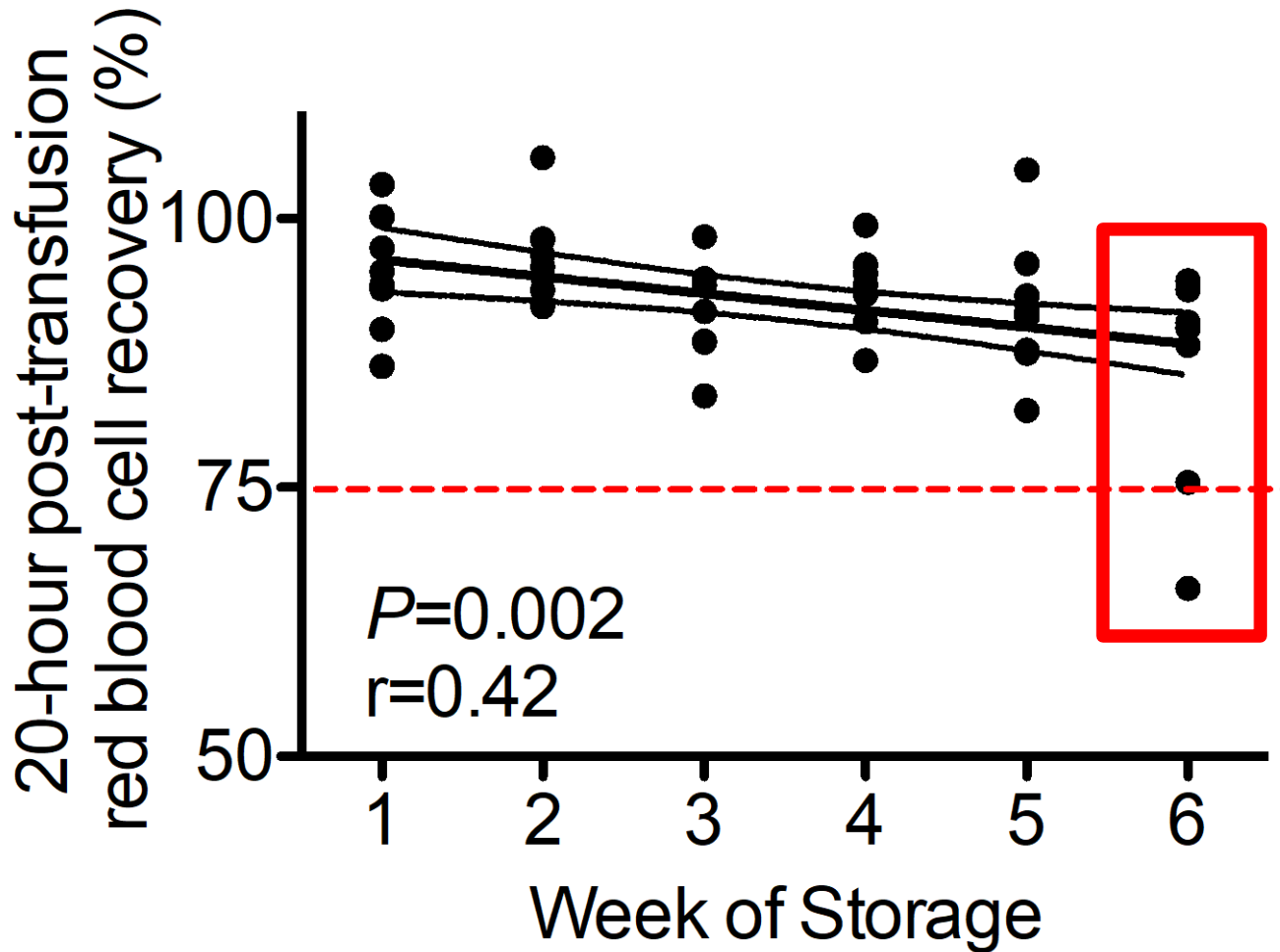
**Similar pattern with serum iron, NTBI, etc.**

**No evidence of intravascular hemolysis**

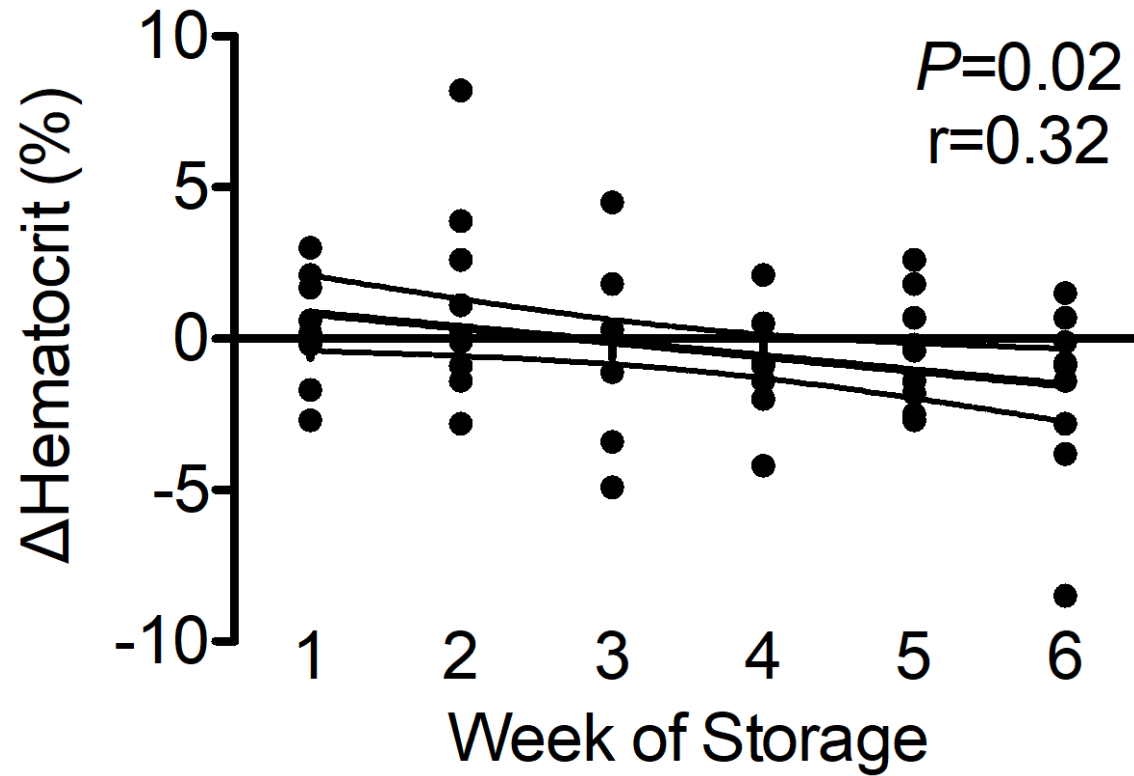
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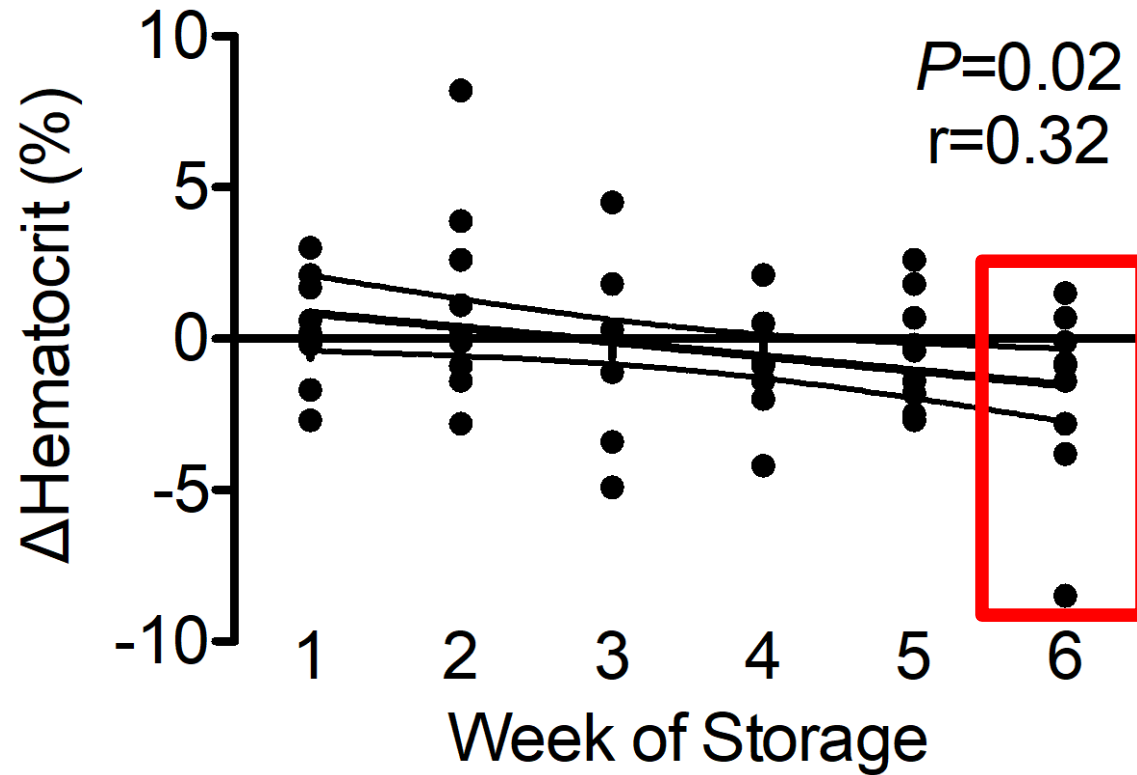
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# When do RBCs “go bad”?



**What donor characteristics  
influence variation in  
post-transfusion recovery?**

# Story #1:

## Genetics: G6PD deficiency



**Richard O. Francis, M.D., Ph.D.**



# The “RBC storage lesion”

## Final common pathway?

Metabolic dysfunction & oxidative stress →

↓ Deformability

↑ “Eat me” signals

↓ “Don’t eat me” signals

↑ Hemolysis *in vitro*

↑ RBC clearance *in vivo*

Intravascular and extravascular hemolysis

# Inherited Hemolytic Anemias

## Final common pathway

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Intravascular and extravascular hemolysis

**What are the consequences of the clearance of stored RBCs?**

**RBC storage lesion *in vitro***



**Decreased RBC recovery *in vivo***

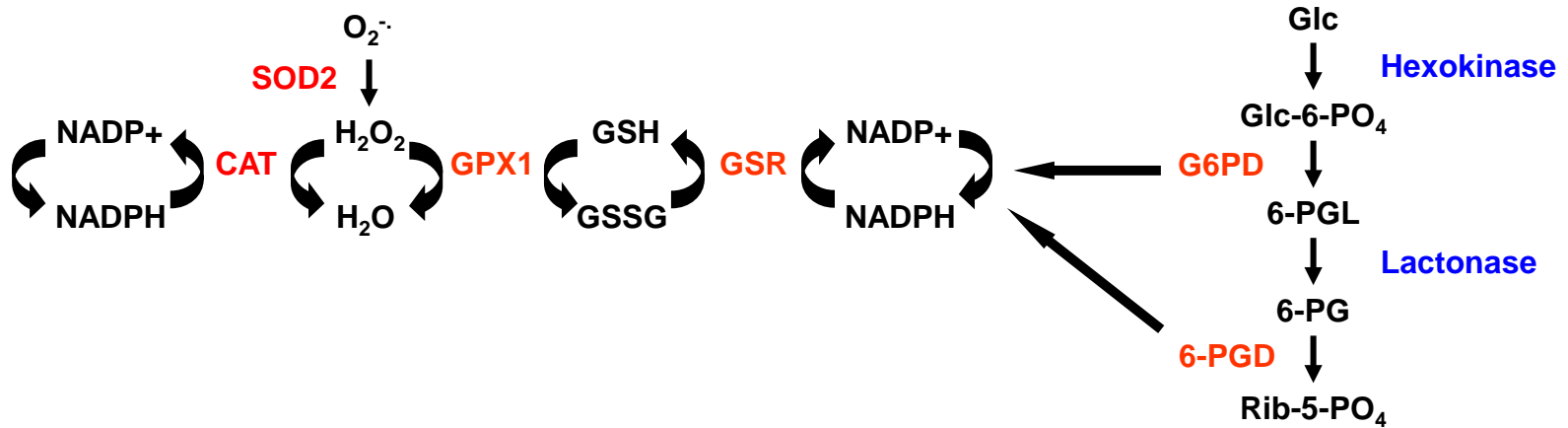
**What are the consequences of the clearance of stored RBCs?**

**Insufficient protection against oxidative stress *in vitro***

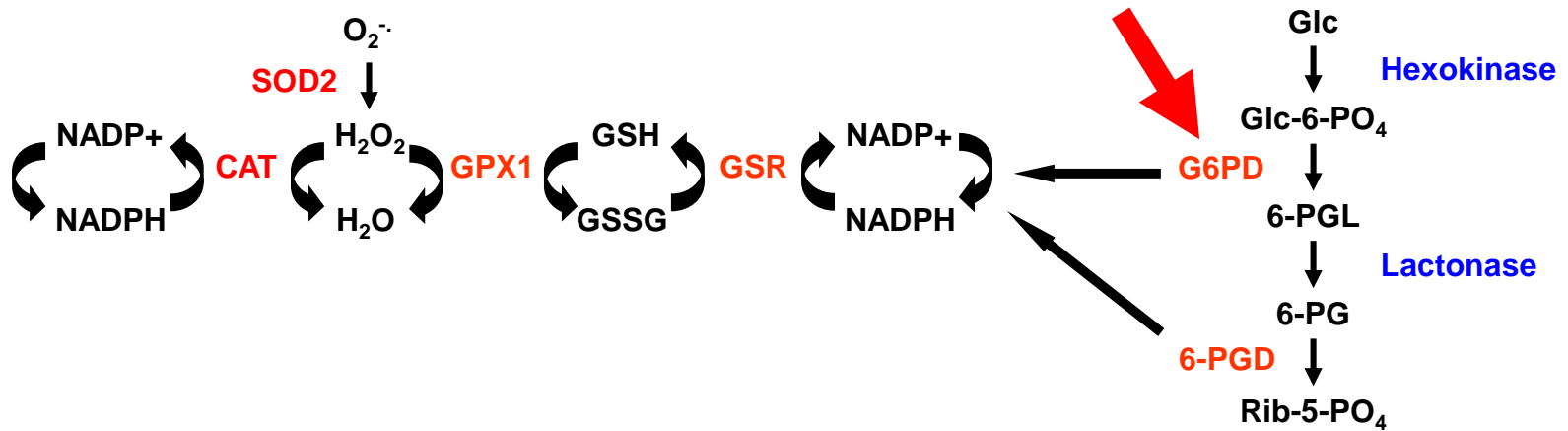


**Decreased RBC recovery *in vivo***

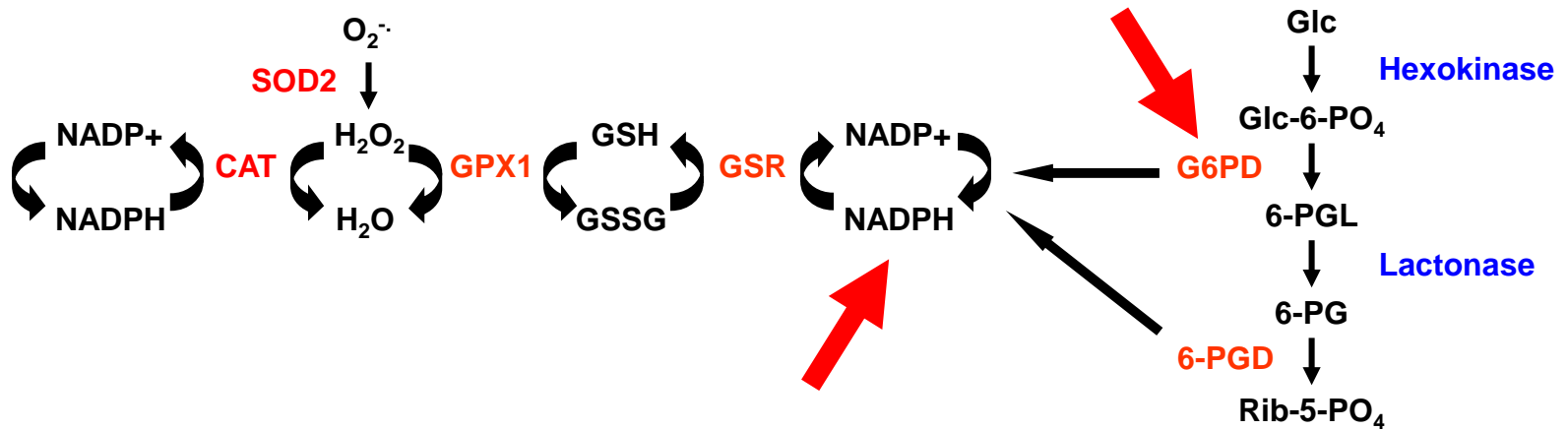
# The case for G6PD: Homeostasis



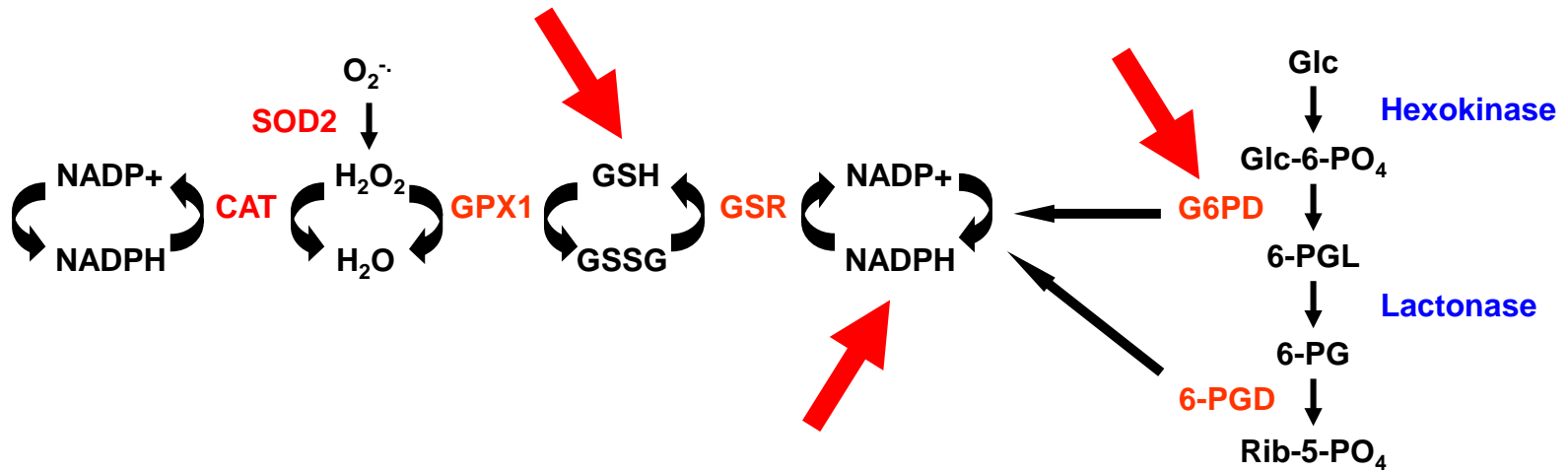
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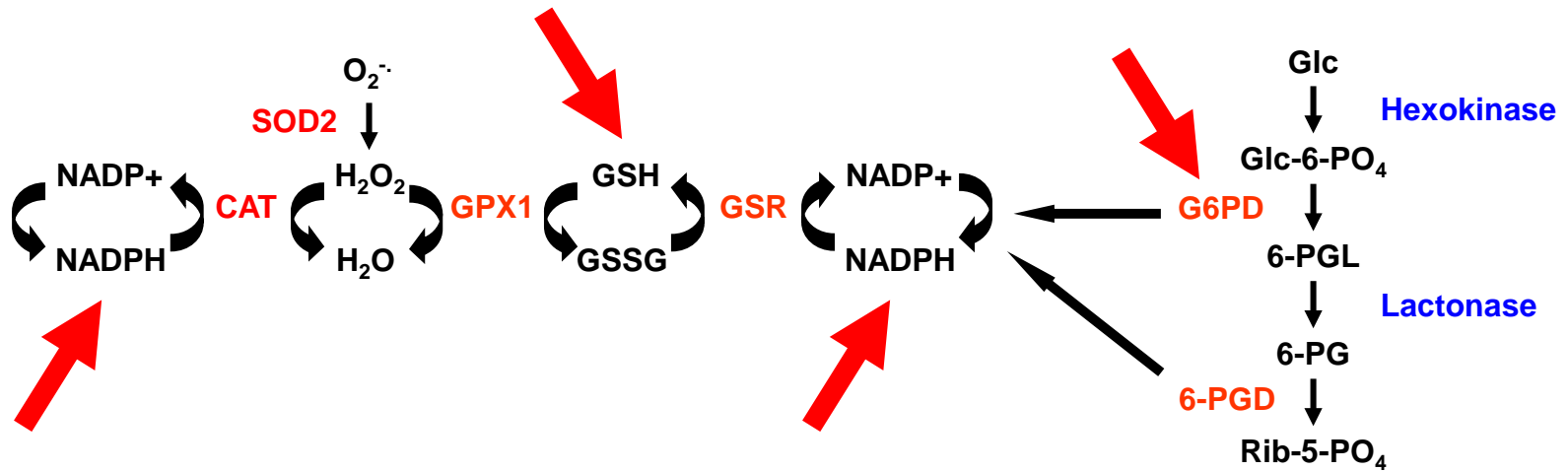


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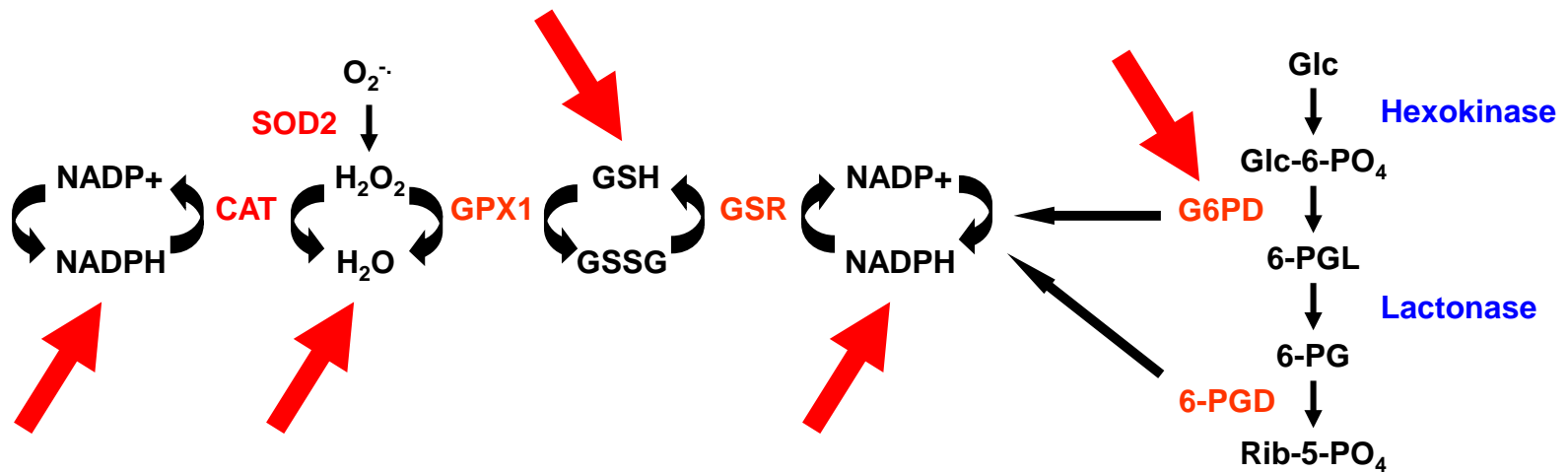




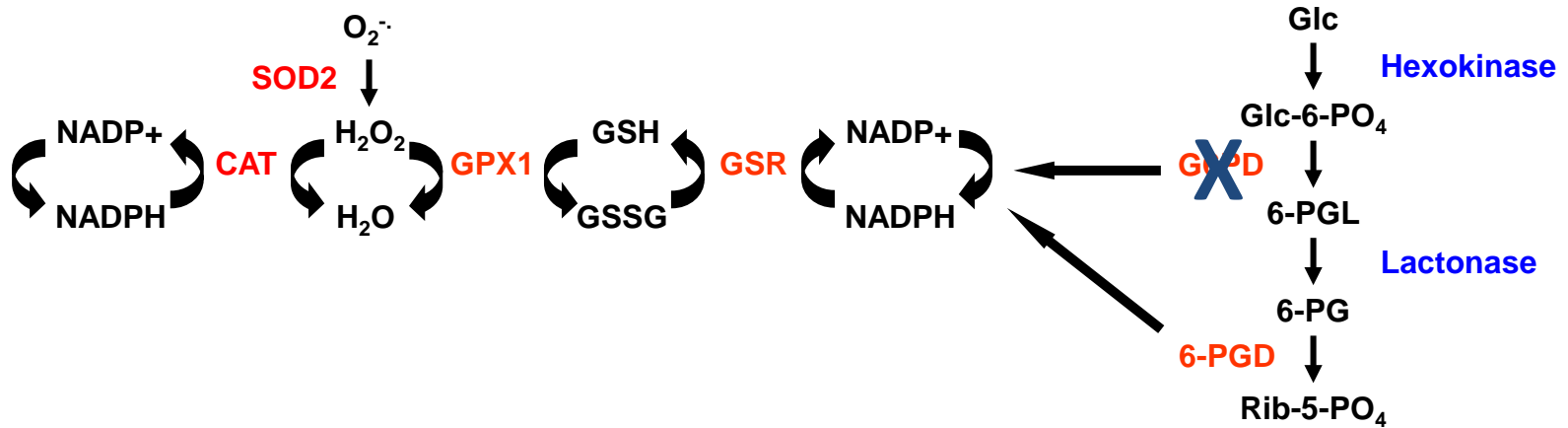
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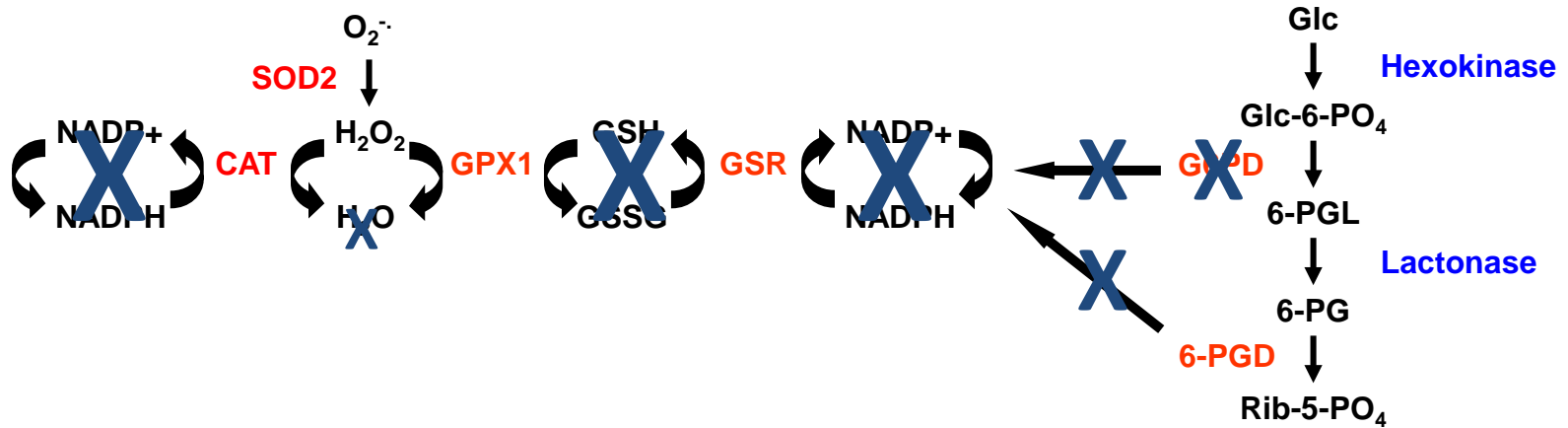
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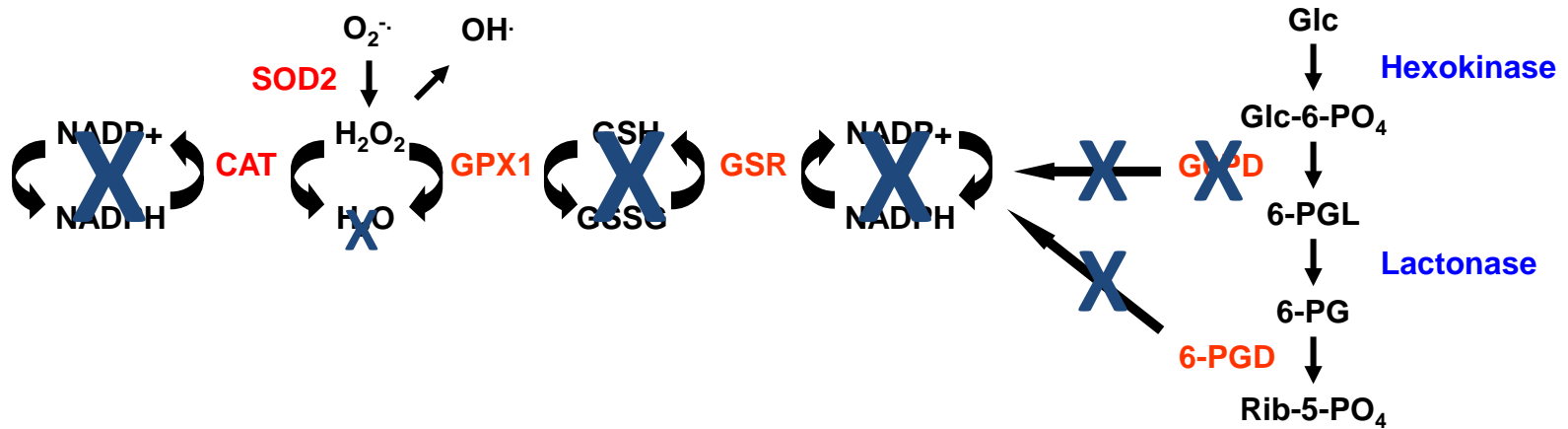
# The case for G6PD: G6PD-deficiency



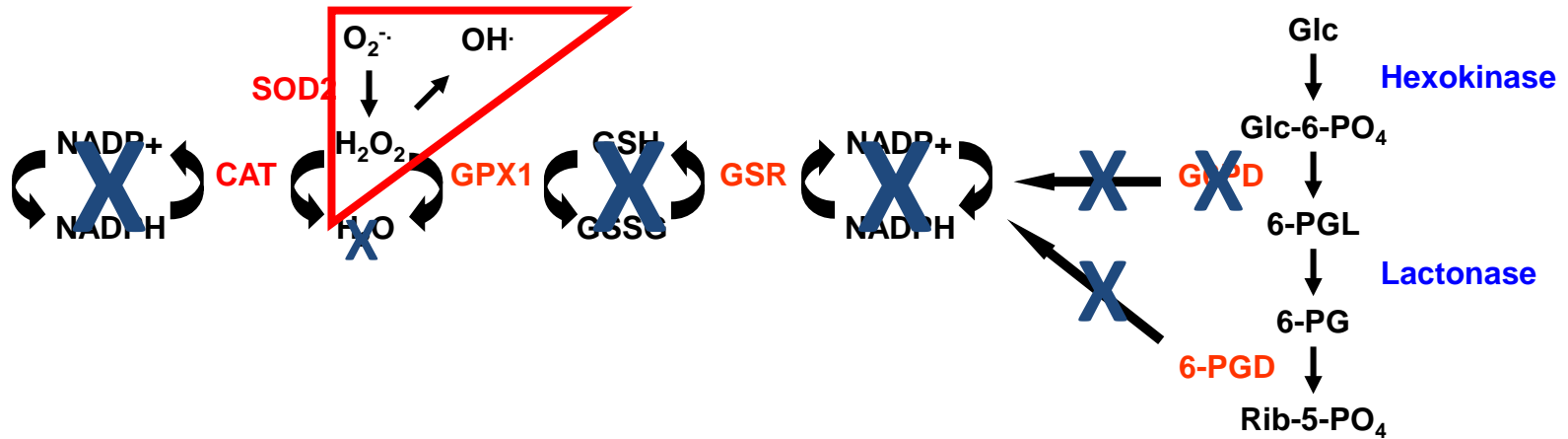
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# **The case for G6PD: G6PD-deficiency**

**Unrelieved oxidative stress:**

**RBC structural damage →**

**Intravascular hemolysis (hemoglobinemia)**

**Extravascular hemolysis (Bilirubin, serum iron)**

# **The case for G6PD: G6PD-deficiency**

**Most common human enzymopathy**



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**~400 million affected individuals**

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**Genetically-induced enzyme variation:**

**Severely decreased activity**

**Normal activity**

**Increased activity**

# The case for G6PD: G6PD-deficiency

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**~400 million affected individuals**

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# The case for G6PD: G6PD-deficiency

Most common human enzymopathy

~400 million affected individuals

Genetically-induced enzyme variation:

Severely decreased activity: **poor storers?**

Normal activity

Increased activity: **super storers?**

# The case for G6PD: G6PD-deficiency



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**Prevalence of G6PD-deficiency in normal donors  
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**Random donors: 0.3%**

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# The case for G6PD: G6PD-deficiency

**Prevalence of G6PD-deficiency in normal donors  
at our hospital:**

**Random donors: 0.3%**

**$R_0R_0/R_0r$  donors: 12.3%**

**Exchange Transfusions for Sickle Cell Disease**

# Study Design

**Volunteers:**

**G6PD-deficient**

**Matched controls**

**Donate 1 unit:**

**Pre-storage leukoreduced**

**Store for 40-42 days in AS-3**

**24h <sup>51</sup>-Cr post-transfusion recovery**

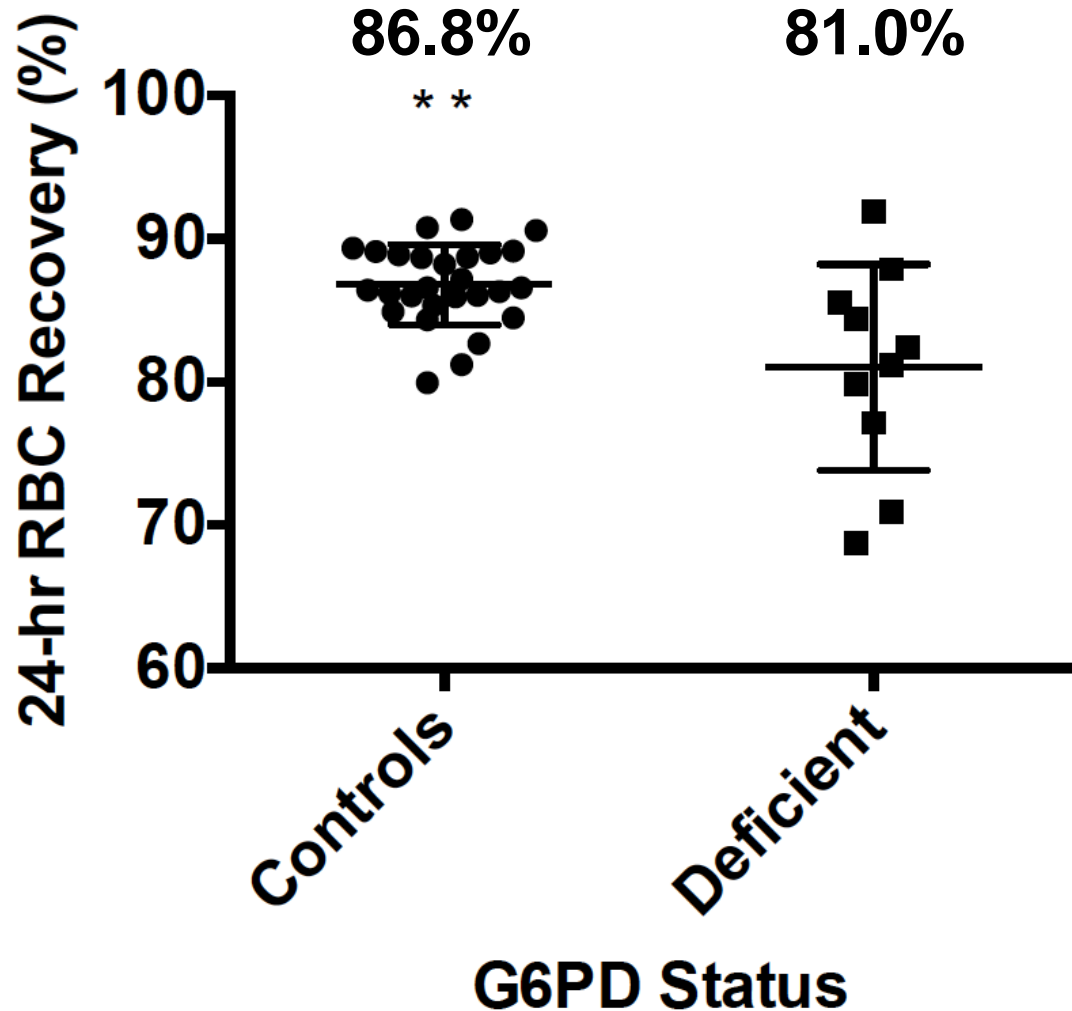
# Study Design

**Enroll: 10 G6PD-deficient + 30 controls**

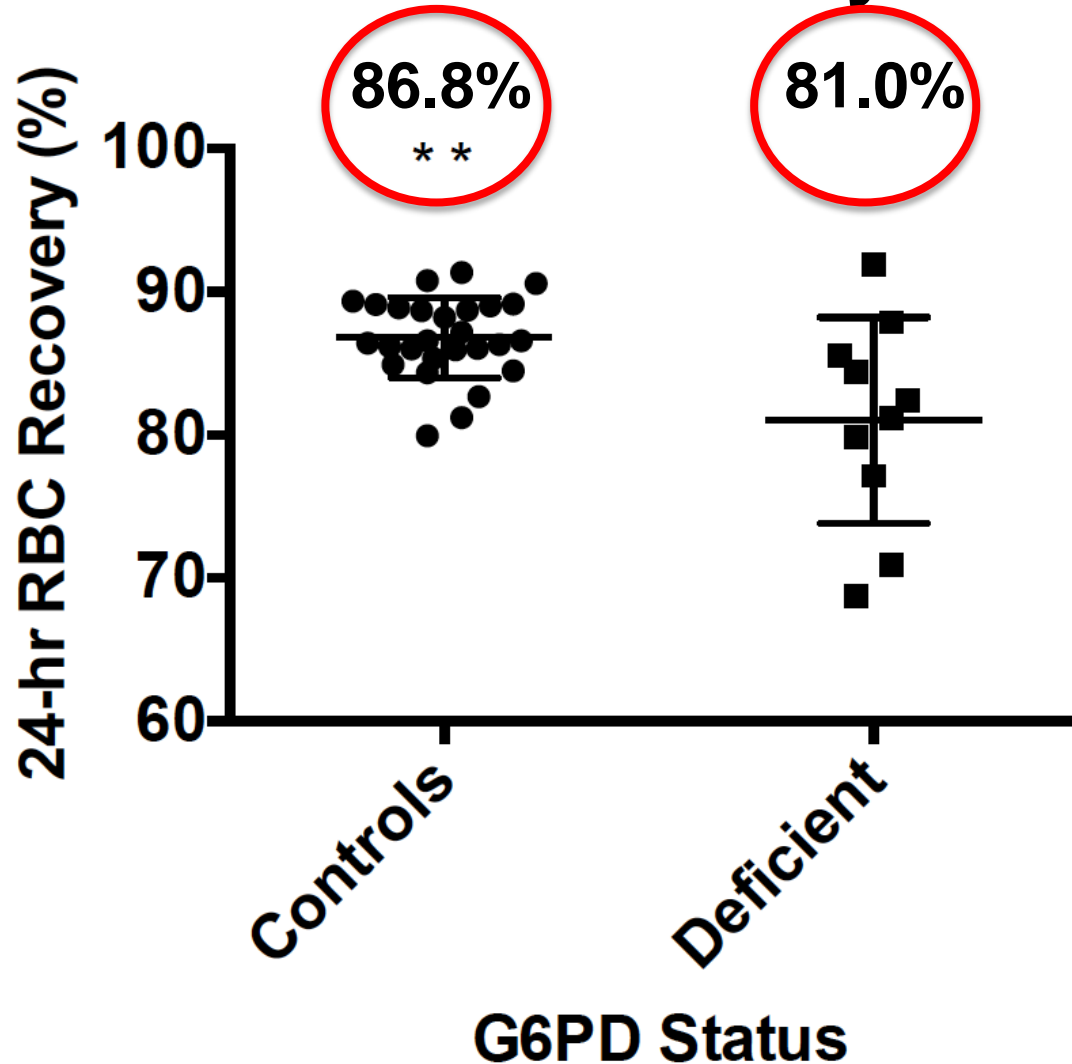
**G6PD-deficient variants: 9 African, 1 Mediterranean**

**None with hemoglobin variant or thalassemia trait**

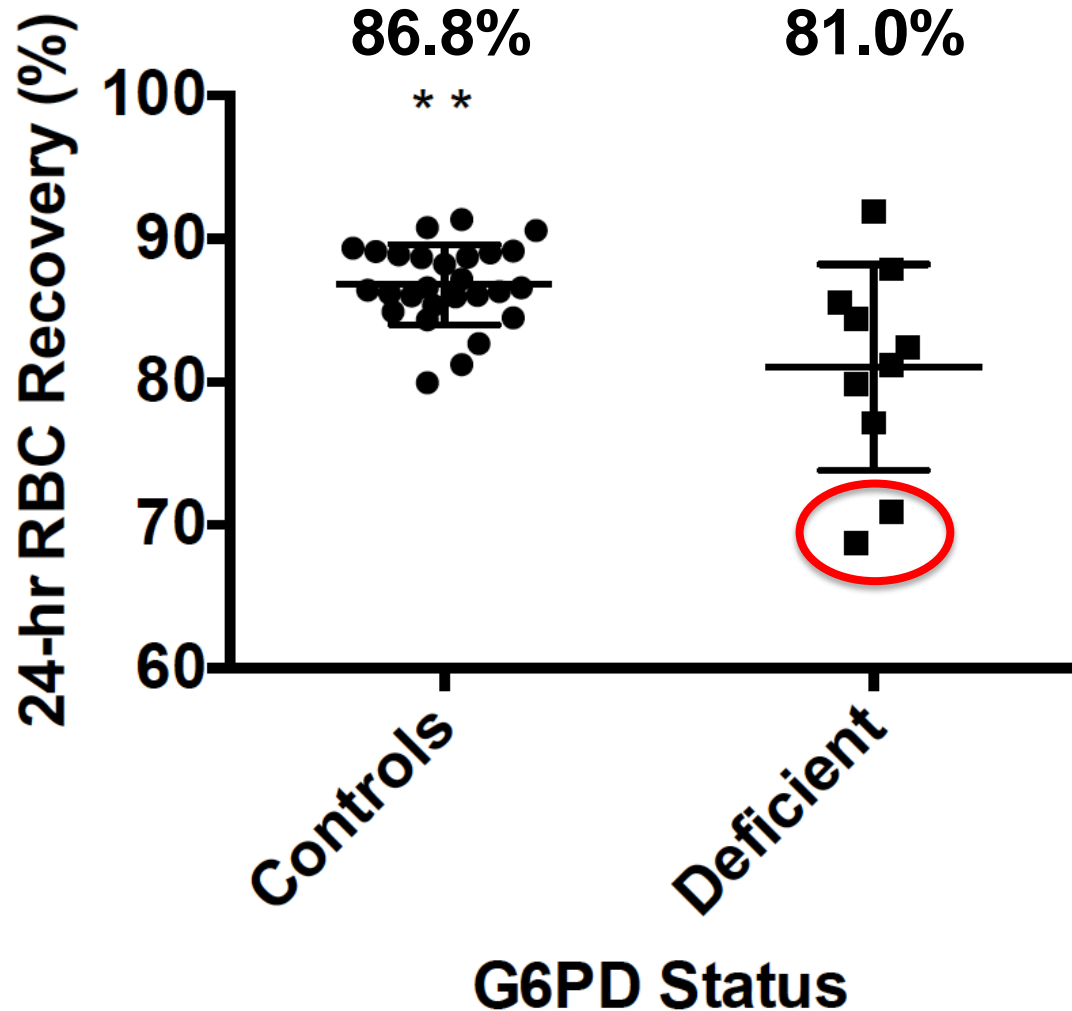
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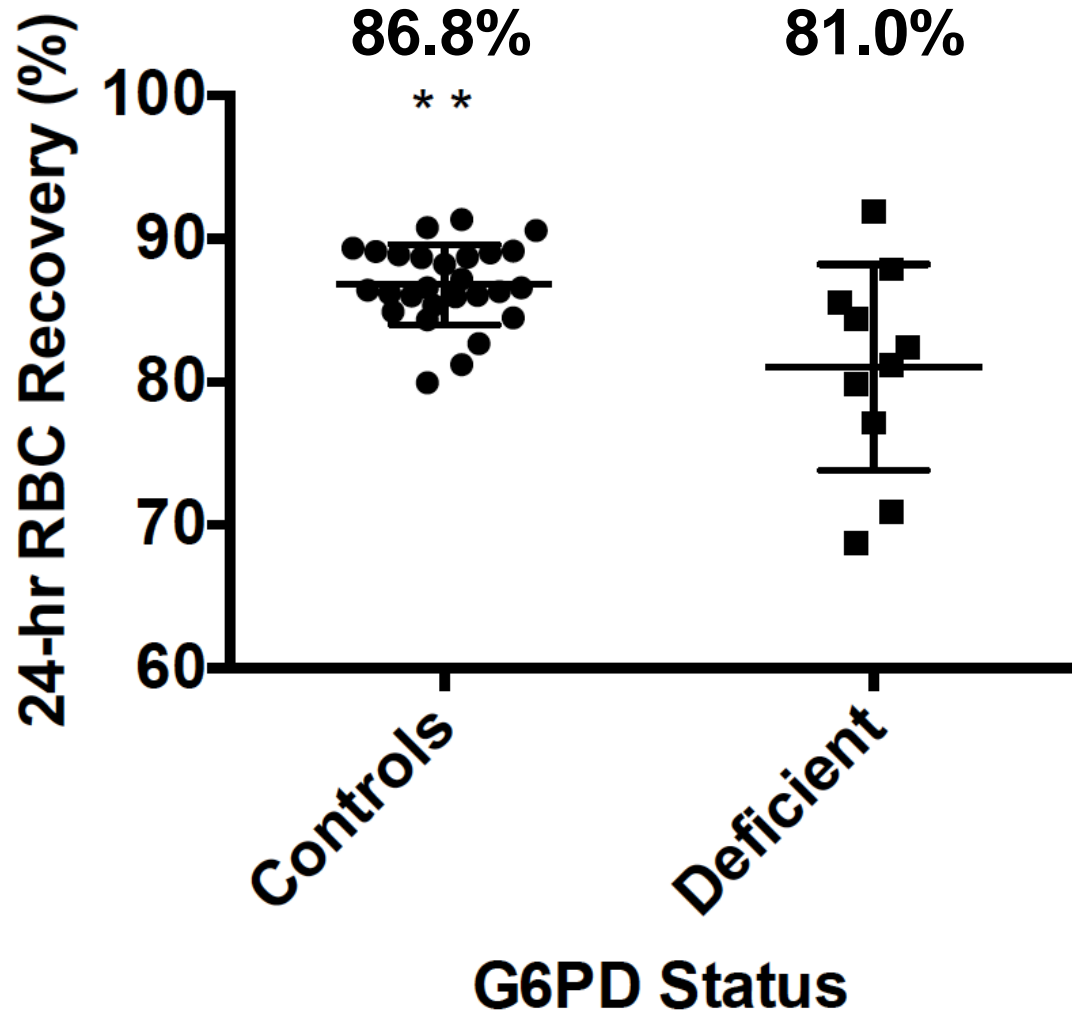


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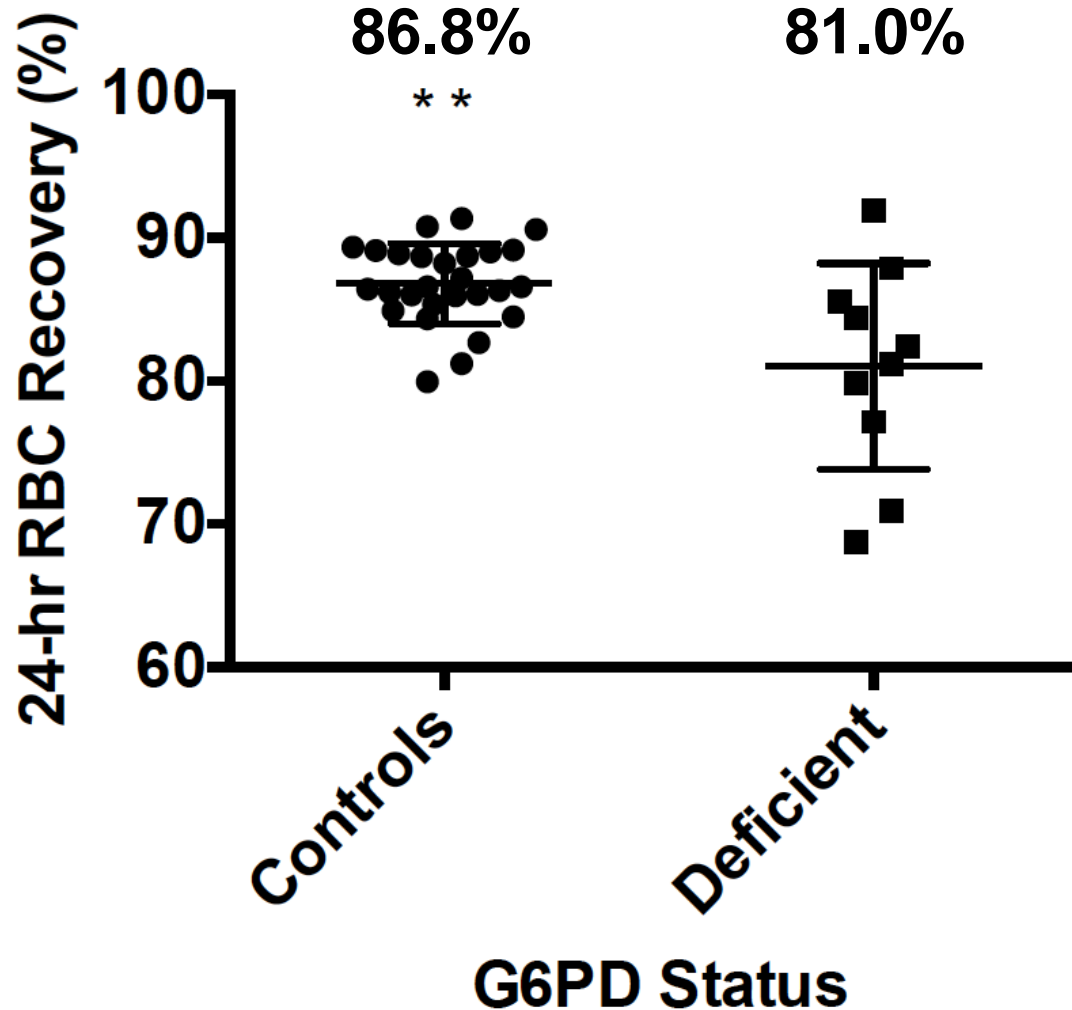
“FDA failures” in  
G6PD-deficient  
group

# The case for G6PD: G6PD-deficiency



No correlation  
between PTR  
and G6PD  
enzyme activity  
within groups

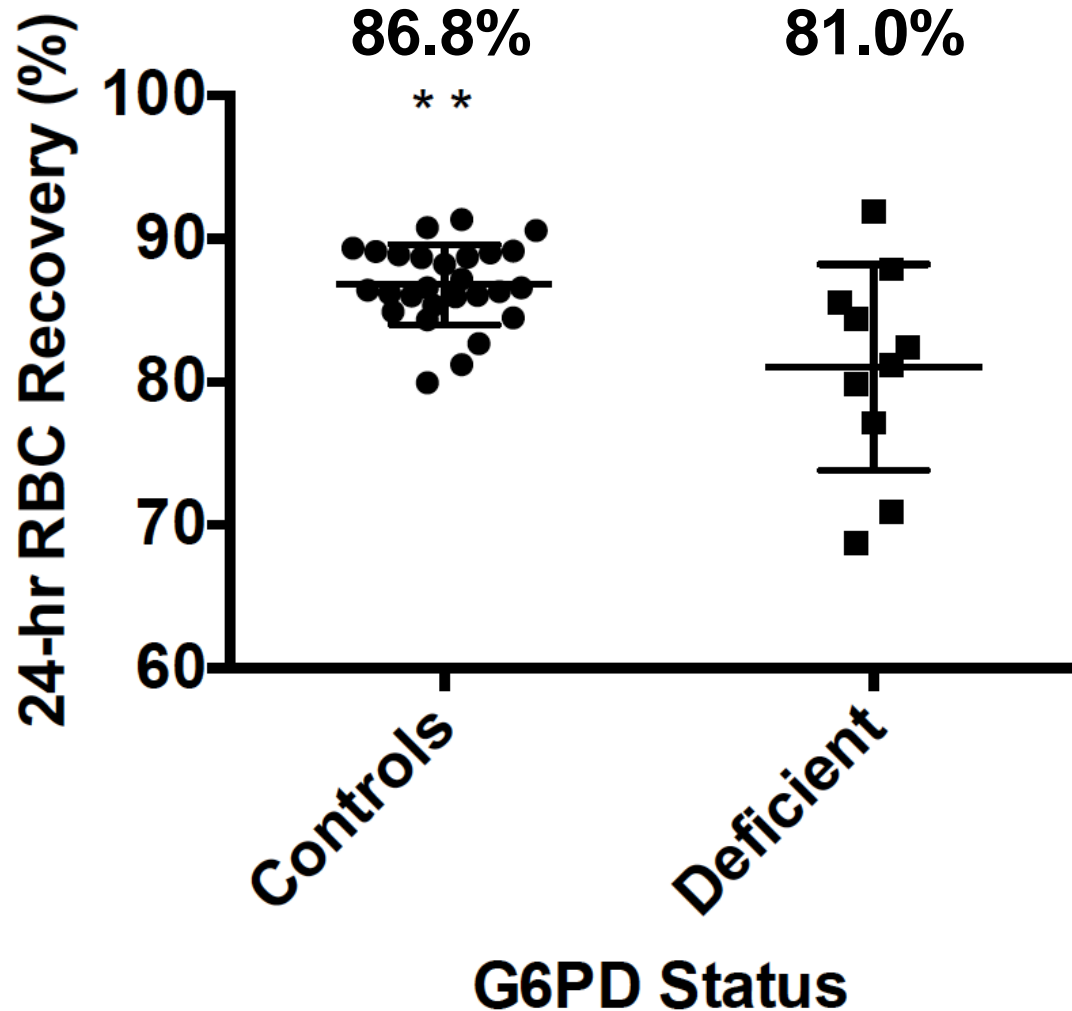
# The case for G6PD: G6PD-deficiency



No difference  
in hemolysis  
“in the bag” at  
outdate



# The case for G6PD: G6PD-deficiency



Would 24-hour post-transfusion recoveries be worse in ill recipients?

# Conclusions (G6PD)

**RBCs from G6PD-deficient volunteers have inferior storage quality: ↓5.8% (p=0.001)**

**The intrinsic ability of RBCs to resist oxidative stress affects storage quality**

**Should we test donors to provide better products?**

**Should we counsel G6PD-deficient donors?**

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**Blood**

---

**Donor**

---

**Selection**

---

**Guidelines on Assessing  
Donor Suitability for  
Blood Donation**

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## Recommendations

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- Policies for the assessment of prospective donors should be developed by BTS in regions where there is a high incidence of enzymopathies and inherited red cell membrane defects

### Accept

- Individuals with G6PD deficiency or other inherited red cell membrane defects, without a history of haemolysis; however, their blood is not suitable for intrauterine transfusion, neonatal exchange transfusion or for patients with G6PD deficiency

### Defer permanently

- Individuals with G6PD deficiency or inherited red cell membrane defects, with a history of haemolysis
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- 

**Should we counsel G6PD-deficient donors regarding their condition?**

# Story #2:

## Diet: Iron deficiency



**Eldad Hod, M.D.**



**Gary Brittenham, M.D.**

# The “RBC storage lesion”

## Final common pathway?

Metabolic dysfunction & oxidative stress →

↓ Deformability

↑ “Eat me” signals

↓ “Don’t eat me” signals

↑ Hemolysis *in vitro*

↑ RBC clearance *in vivo*

Intravascular and extravascular hemolysis

**Iron deficiency (without anemia) is very common in blood donors**



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very common in blood donors**

**Iron-deficient erythropoiesis (IDE)**

# Iron deficiency **anemia** affects RBC lifespan & transfusion recovery

↓ Resistance to oxidative stress

↑ Oxidative damage

↓ Resistance to low pH

↑ Phosphatidylserine exposure

↓ Deformability

↑ Splenic clearance

# Iron deficiency **anemia** affects RBC lifespan & transfusion recovery

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**Is this relevant in IDE?**

# Mouse model

Weanling, male C57BL/6 mice:

1. Control diet: 45 ppm of iron (normal)
2. Iron-deficient diet: 0-4 ppm of iron (IDE)
3. Iron-deficient diet + weekly phlebotomy (IDA)

# Mouse model

Weanling, male C57BL/6 mice:

1. Control diet: 45 ppm of iron (normal)
2. Iron-deficient diet: 0-4 ppm of iron (IDE)
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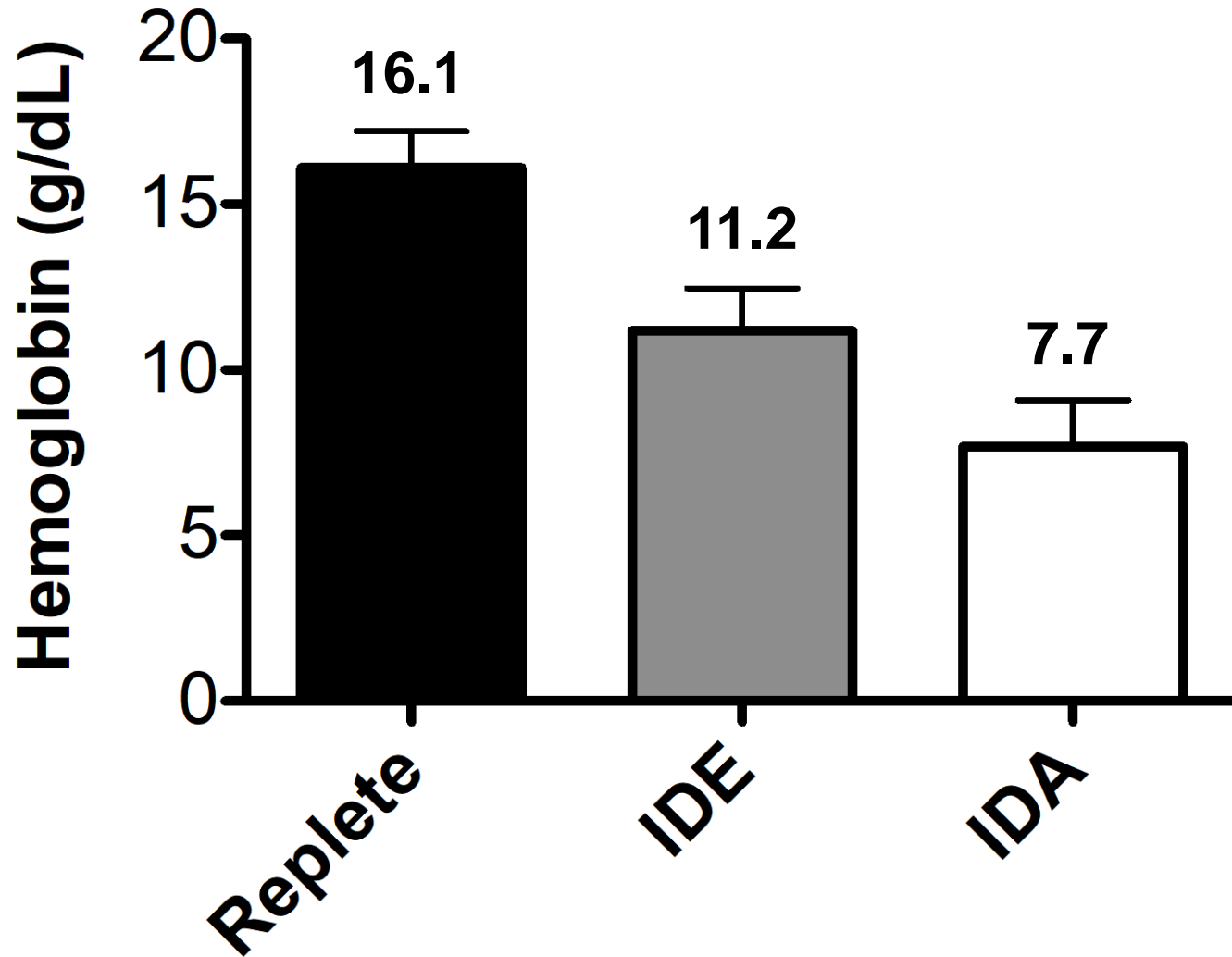
**At 10 weeks of age:**

**Exsanguinated, pooled, leukoreduced, packed,  
stored in CPDA-1 for 12 days**

**Transfused into GFP-transgenic recipients**

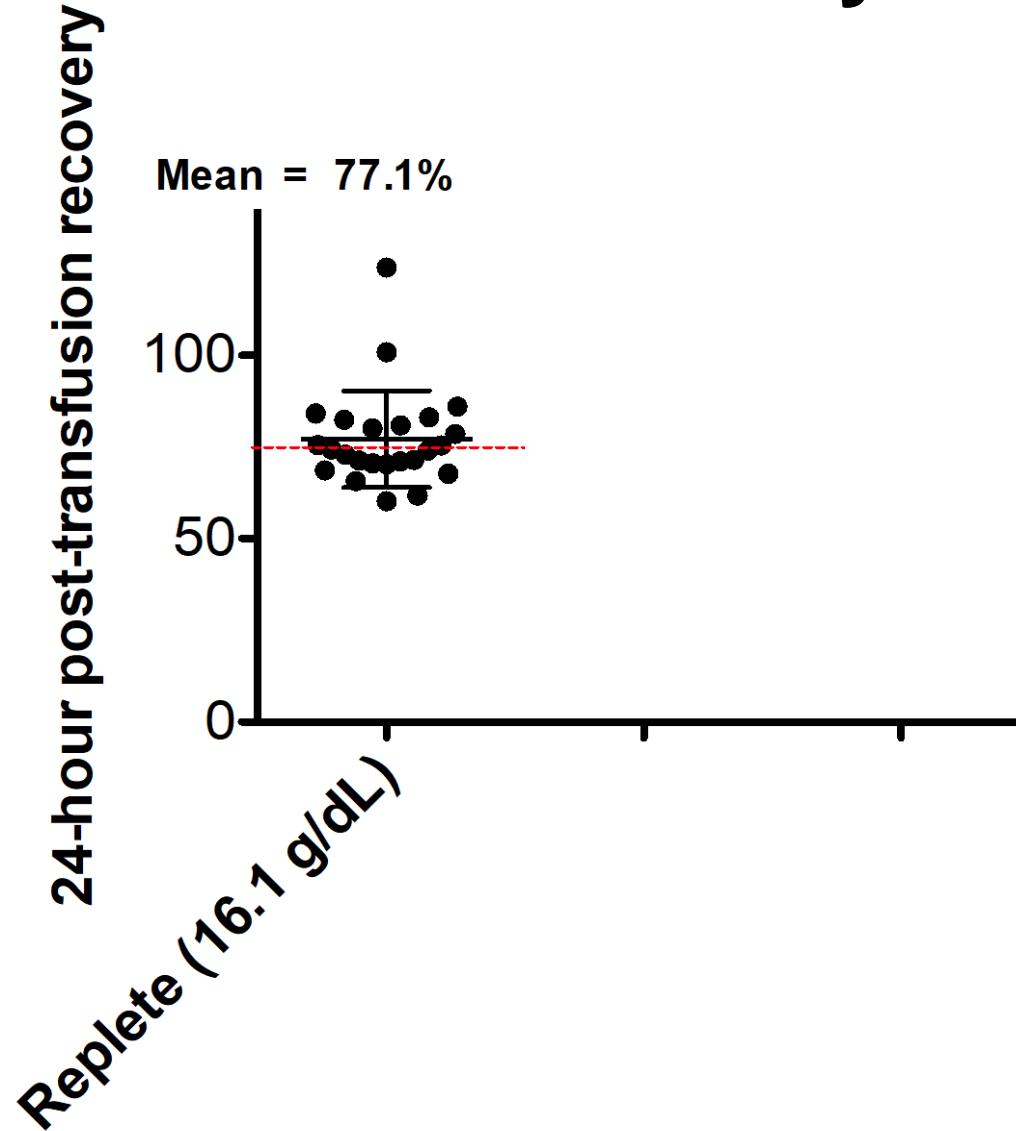
**24-hour post-transfusion recovery by flow cytometry**

# Results

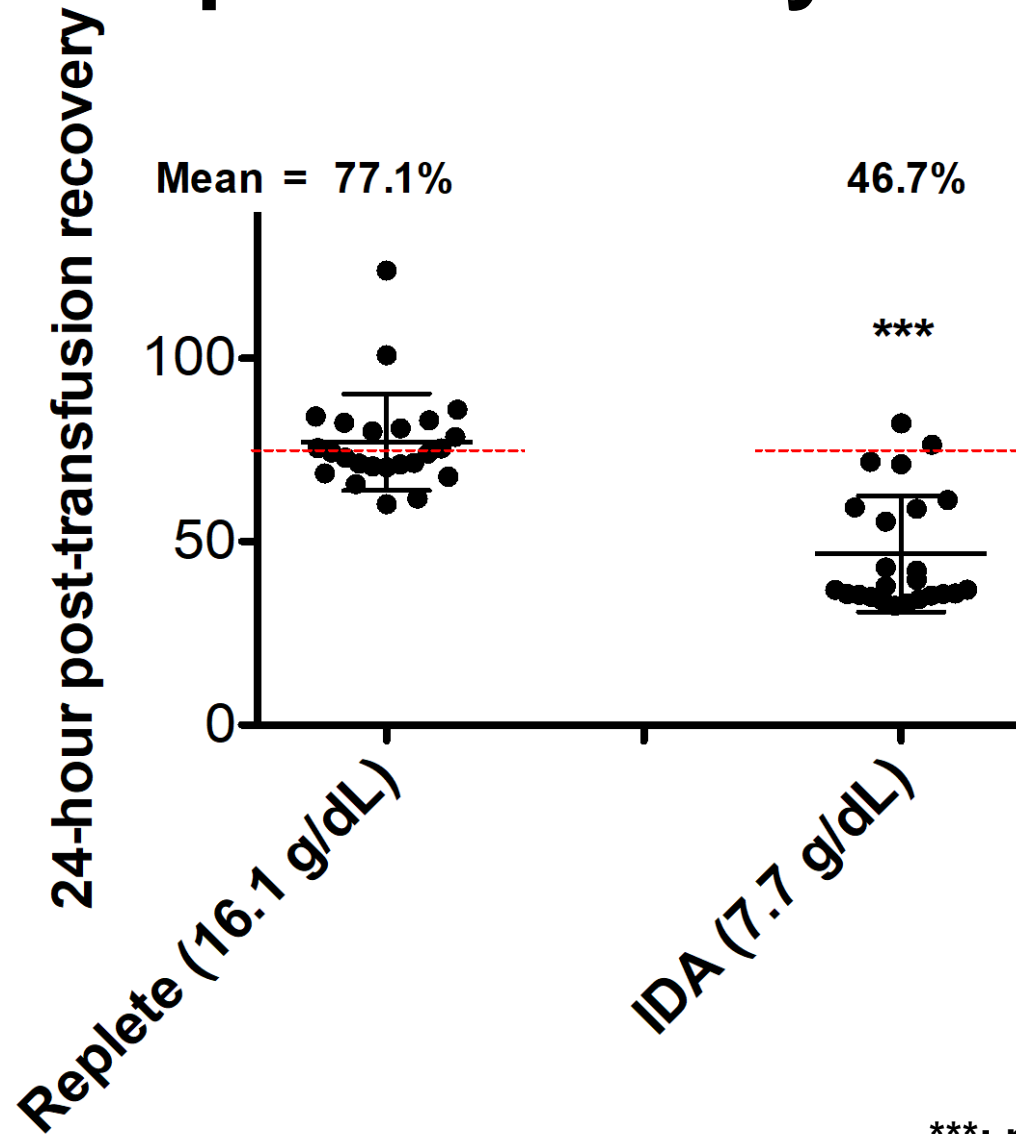


All comparisons:  $p < 0.001$

# RBCs from normal donors had normal recovery



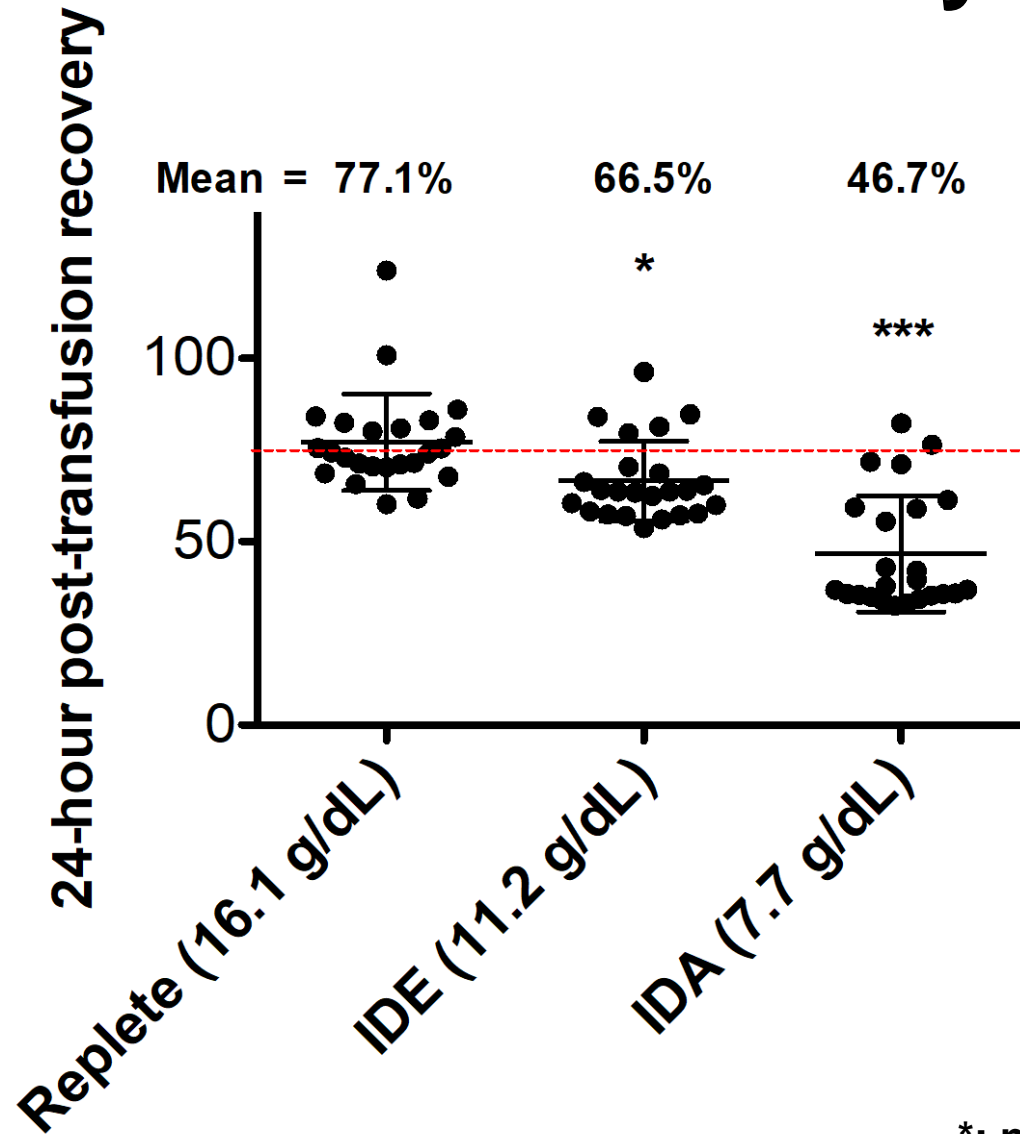
# RBCs from donors with IDA had poor recovery



\*\*\*: p<0.001



# RBCs from donors with **IDE** had sub-normal recovery



**Would this be true for human recipients of RBC transfusions from donors with IDE?**

# Would this be true for human recipients of RBC transfusions from donors with IDE?

ClinicalTrials.gov

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Trial record 3 of 5 for: eldad hod

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## Donor Iron Deficiency Study - Red Blood Cells From Iron-deficient Donors: Recovery and Storage Quality (DIDS)

**This study is enrolling participants by invitation only.**

**Sponsor:**

Columbia University

**Collaborators:**

New York Blood Center  
National Heart, Lung, and Blood Institute (NHLBI)

**Information provided by (Responsible Party):**

Eldad Arie Hod, Columbia University

**ClinicalTrials.gov Identifier:**

NCT02889133

First received: August 25, 2016

Last updated: March 20, 2017

Last verified: March 2017

[History of Changes](#)

[Full Text View](#)

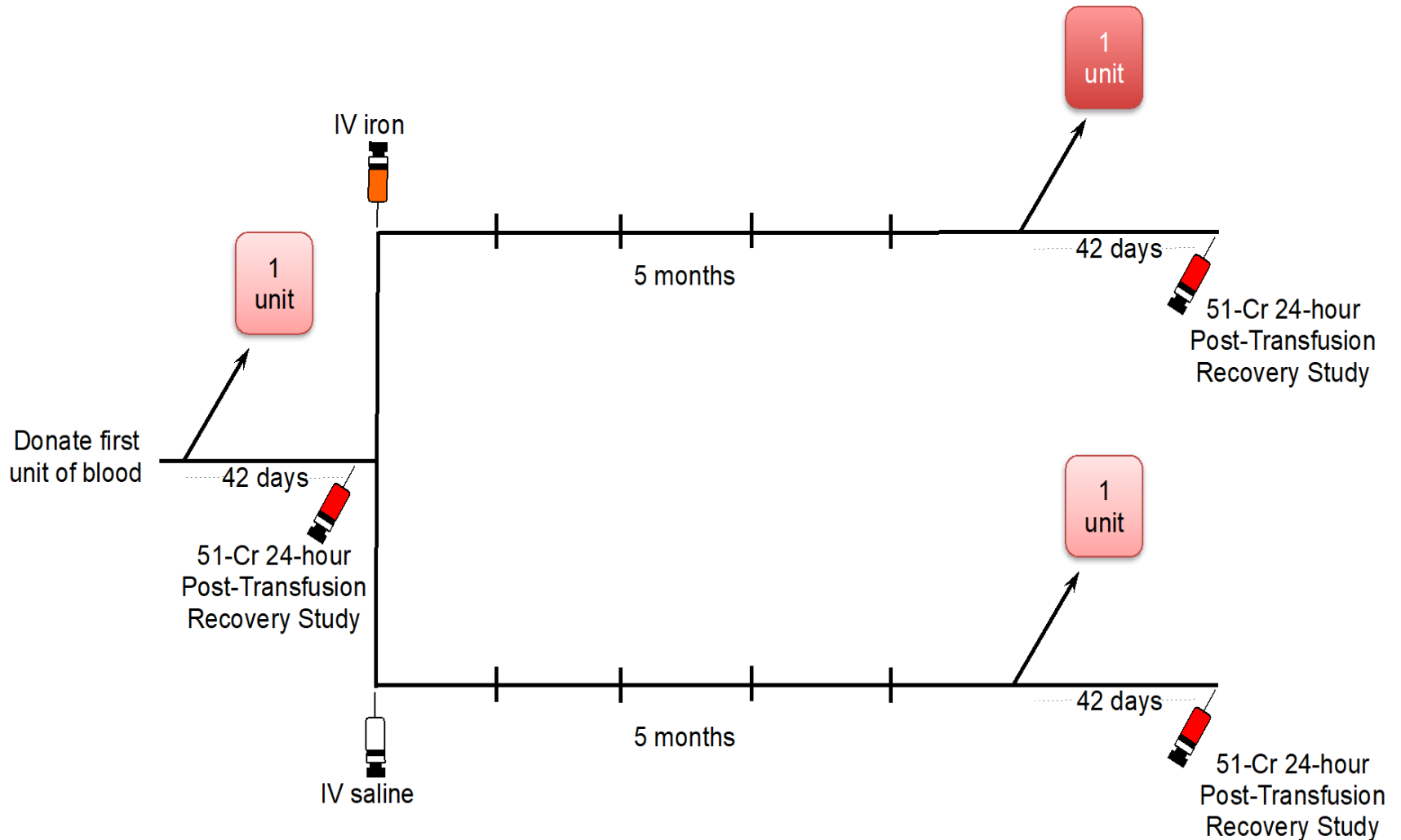
[Tabular View](#)

[No Study Results Posted](#)

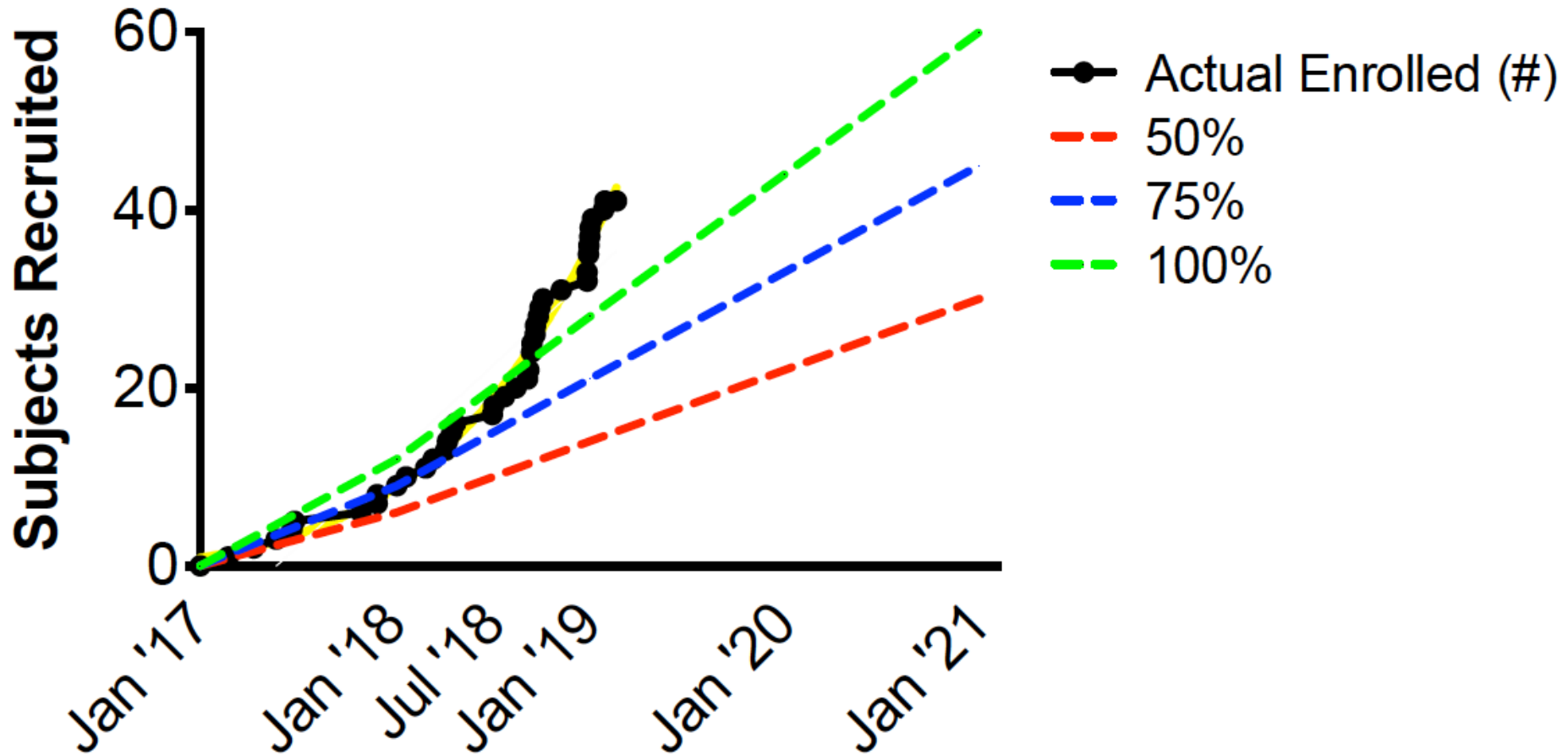
[Disclaimer](#)

[? How to Read a Study Record](#)

# Would this be true for human recipients of RBC transfusions from donors with IDE?



# Would this be true for human recipients of RBC transfusions from donors with IDE?



# **Conclusions (Iron)**

**Should we screen donors for iron-deficient erythropoiesis?**

**Should we collect & transfuse RBCs from such donors?**

**Should we counsel & treat iron-deficient donors?**

# Story #3:

## “Environment”: Lead



**Tiffany Thomas, Ph.D.**

**Lead**



# **Lead**

**Neurotoxicant (synapses, myelin, etc.)**

# **Lead**

**Neurotoxicant (synapses, myelin, etc.)**

**No threshold effect**

# **Lead**

**Neurotoxicant (synapses, myelin, etc.)**

**No threshold effect**

**Young children particularly vulnerable**

# **Lead**

**Neurotoxicant (synapses, myelin, etc.)**

**No threshold effect**

**Young children particularly vulnerable**

**Premature babies are relatively Fe deficient**

# Lead

**Neurotoxicant (synapses, myelin, etc.)**

**No threshold effect**

**Young children particularly vulnerable**

**Premature babies are relatively Fe deficient**

**↑ blood lead level:  $>5 \mu\text{g/dL}$  ( $0.2415 \mu\text{mol/L}$ )**

# Lead

**Neurotoxicant (synapses, myelin, etc.)**

**No threshold effect**

**Young children particularly vulnerable**

**Premature babies are relatively Fe deficient**

**↑ blood lead level:  $>5 \mu\text{g/dL}$  ( $0.2415 \mu\text{mol/L}$ )**

**In whole blood, 75% of lead is in RBCs**

# Lead

**Do donor pRBC units contain high lead levels?**

# Lead

**Do donor pRBC units contain high lead levels?**

**If so, who cares?**



# Lead

## BLOOD DONORS AND BLOOD COLLECTION

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### **A population-based study on blood lead levels in blood donors**

*Gilles Delage,<sup>1</sup> Suzanne Gingras,<sup>2</sup> and Marc Rhainds<sup>2,3</sup>*

**TRANSFUSION 2015;55;2633–2640**

# Lead

## BLOOD DONORS AND BLOOD COLLECTION

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**“Toxic” levels in ~10% of their donor population**

# Lead

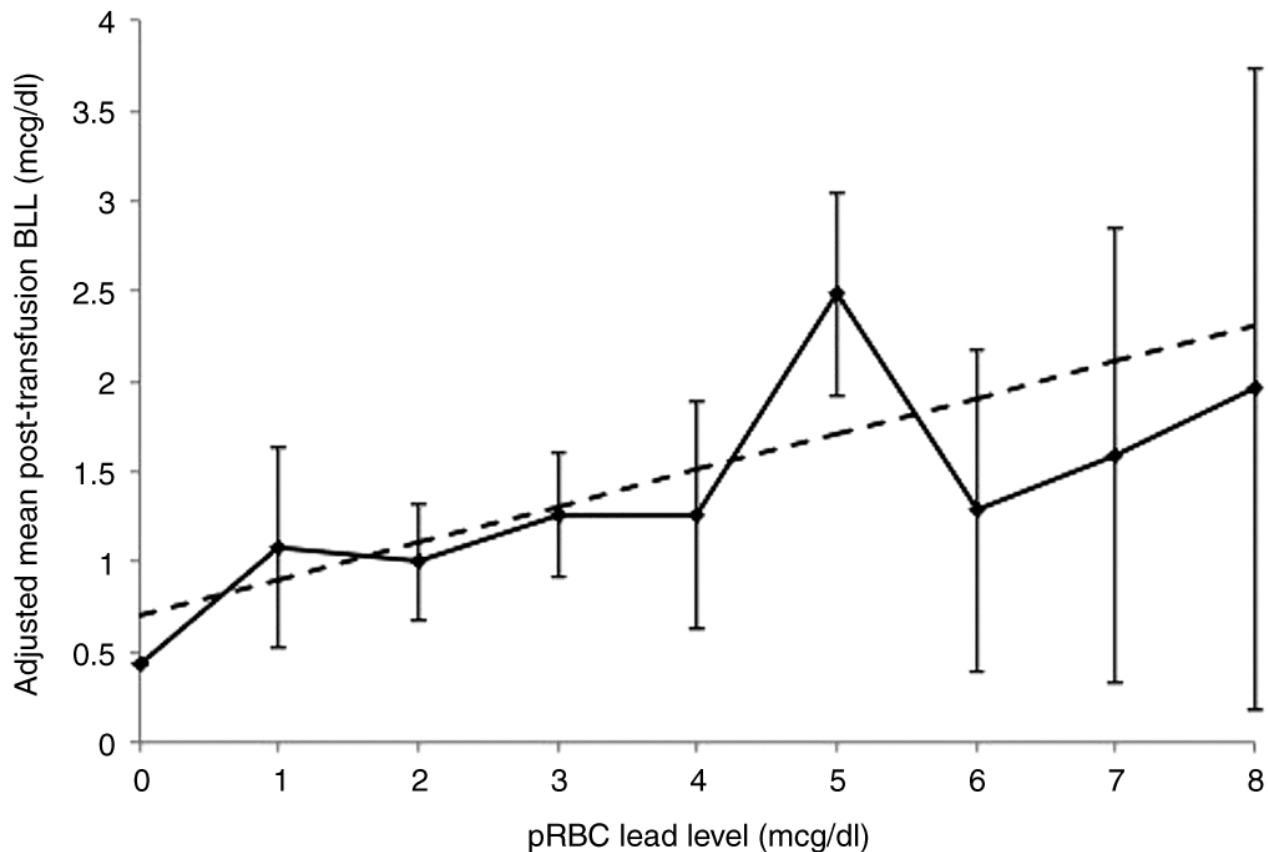
## **Lead exposure in preterm infants receiving red blood cell transfusions**

Hijab Zubairi<sup>1</sup>, Paul Visintainer<sup>2,3</sup>, Jennie Fleming<sup>1</sup>, Matthew Richardson<sup>1,3</sup> and Rachana Singh<sup>1,3</sup>

Pediatric RESEARCH

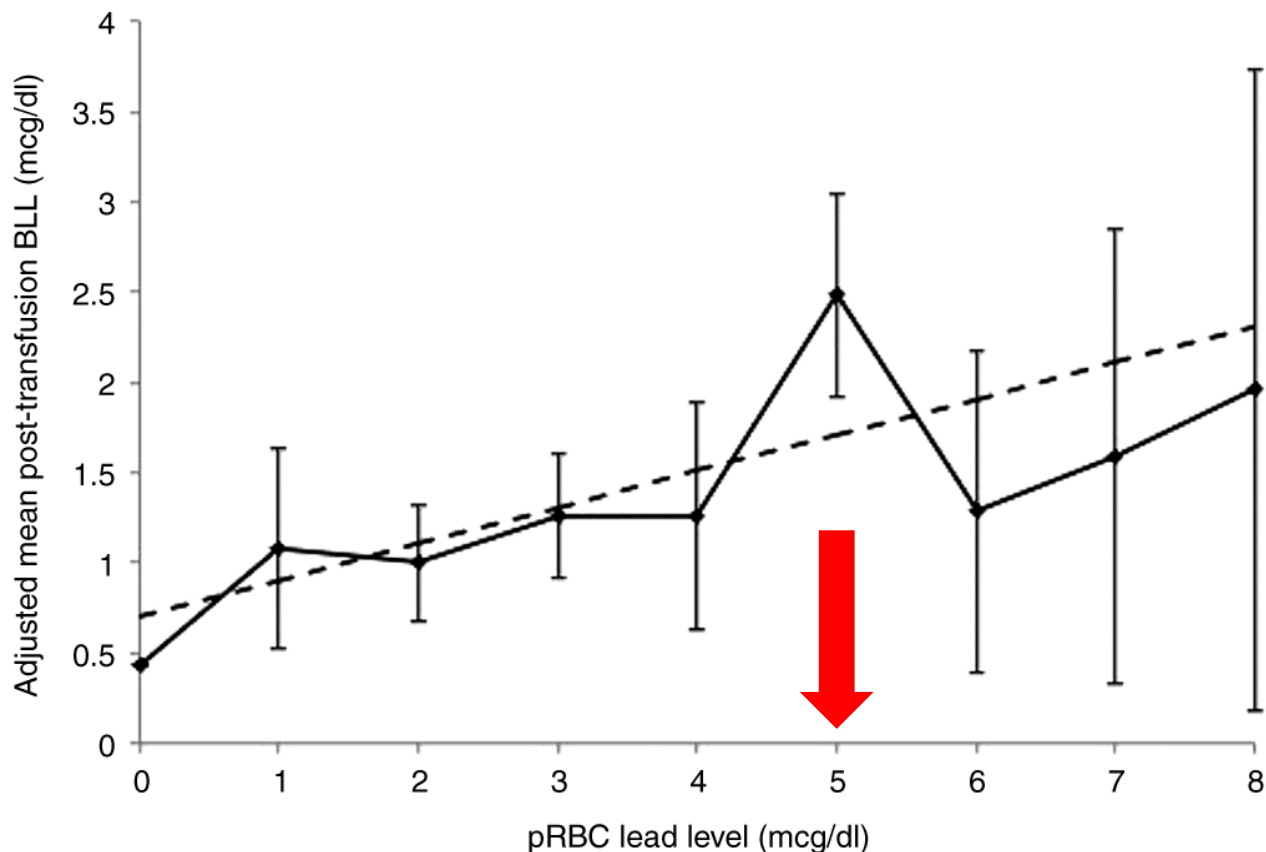
Volume 77 | Number 6 | June 2015

# Lead



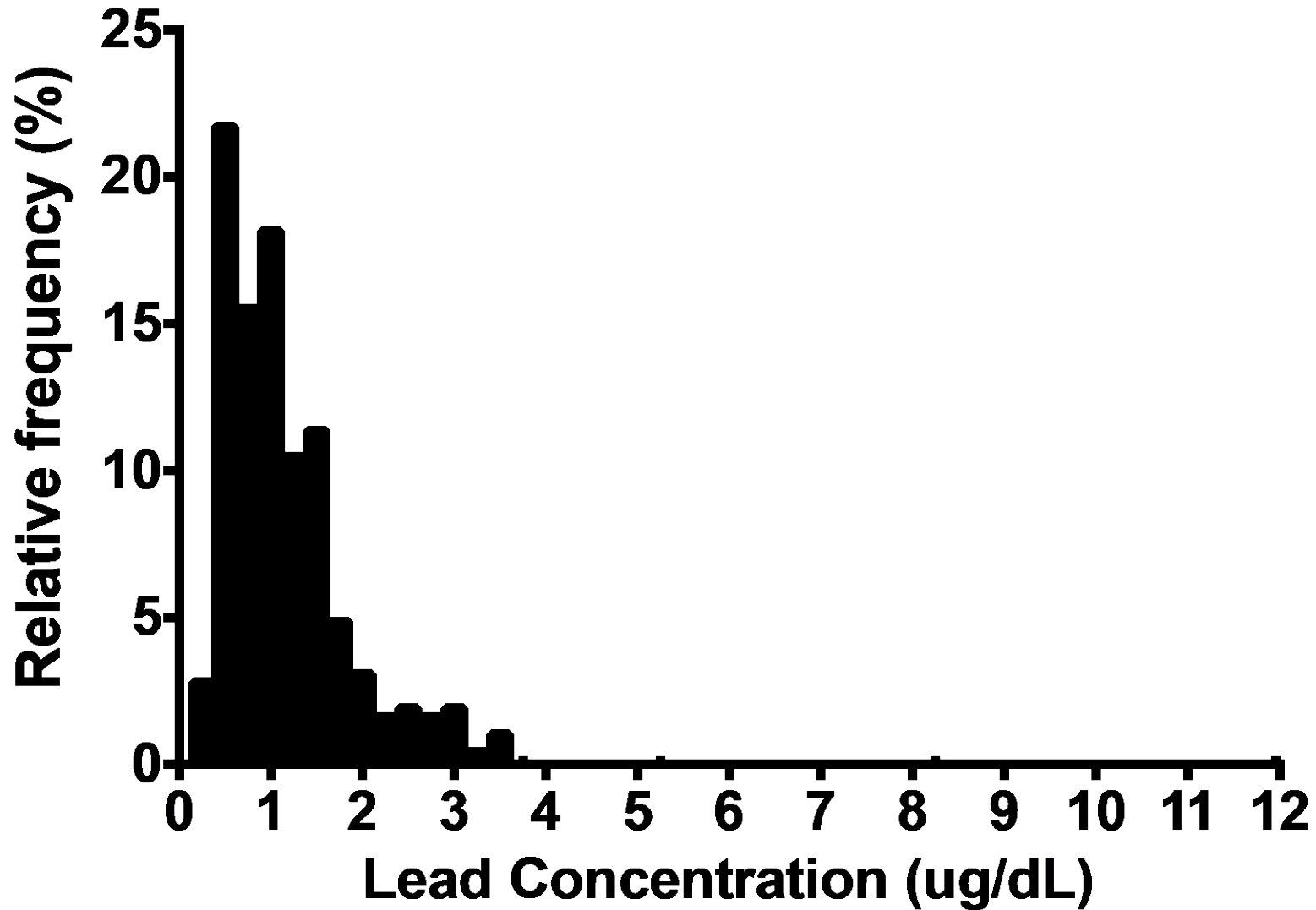
**Figure 1.** Adjusted mean (and 95% CI) post-transfusion BLL by pRBC lead level. For each 1 mcg/dl increase in transfused pRBC lead level, infant post-transfusion BLL increased by 0.20 mcg/dl (95% CI: 0.07 mcg/dl, 0.33 mcg/dl; test of linear trend:  $P = 0.002$ ). The means are adjusted for gestational age, birth weight, and pretransfusion blood lead level.

# Lead

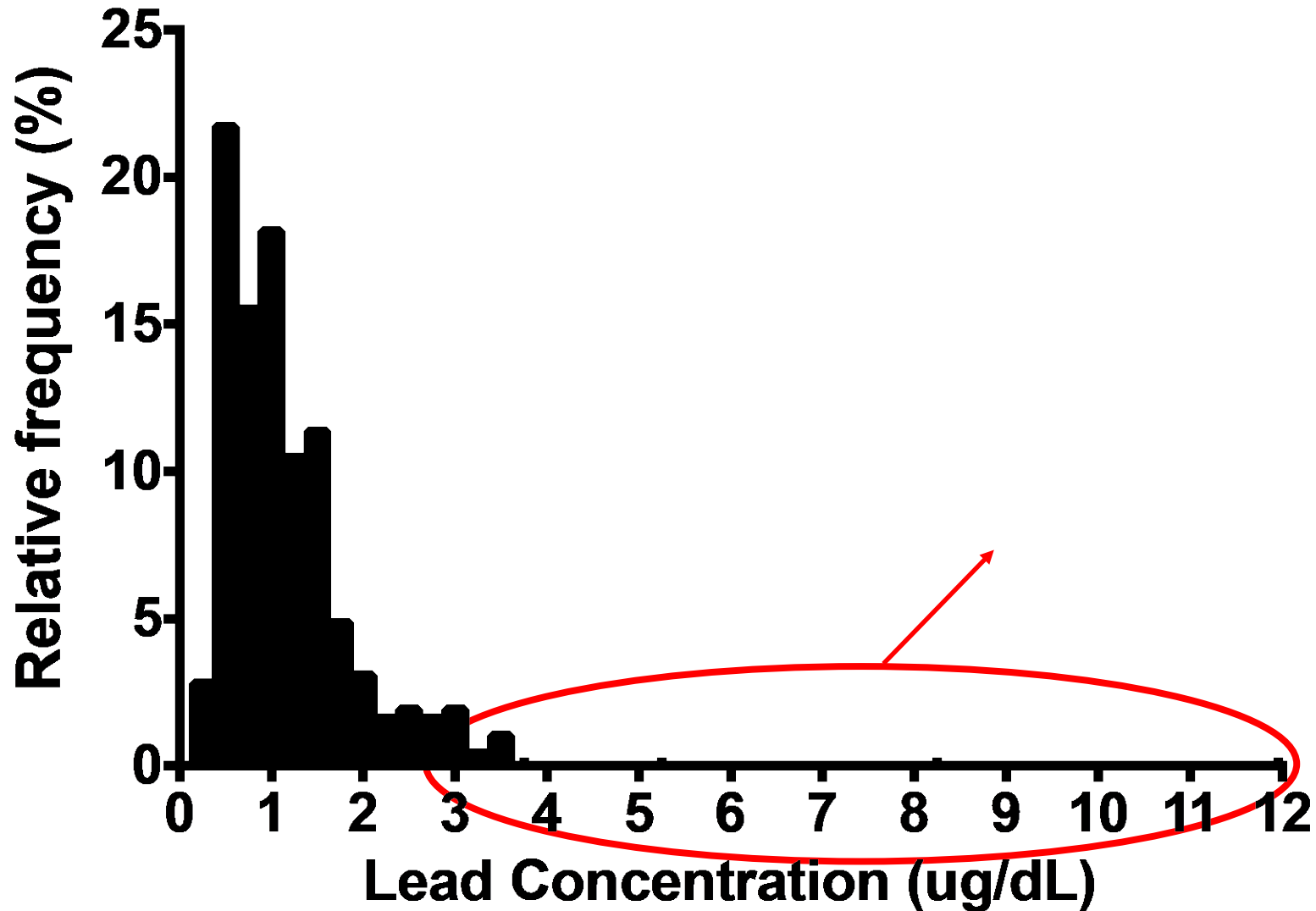


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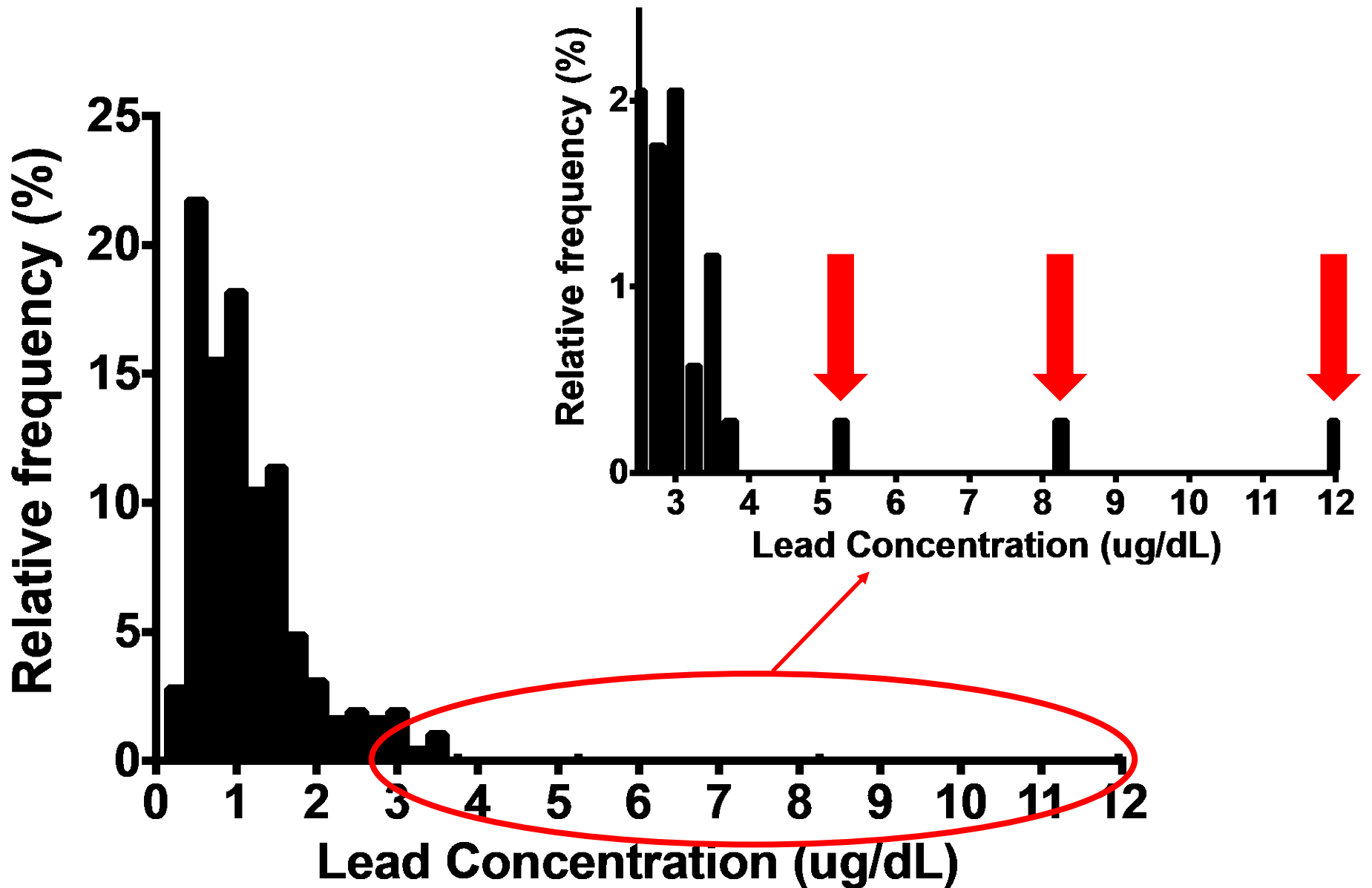
# Our data



# Our data



# Our data





# **Conclusions: Lead**

**Should we screen units destined for premature infants?**

**Should we inform donors of their lead “toxicity”?**

# **Future Directions**

**Making better products: Ideal RBC unit**

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“Equivalent to fresh”**

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**No plasma**

**Avoid allergic NHTRs, anaphylactic NHTRs, and TRALI**

**Lacking clinically-significant RBC blood group antigens**

**Avoid RBC alloimmunization and HTRs**

# Future Directions

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**If so, what ethical obligations do we have to protect and inform our donors?**



**Thank you**