



The efficient use of RhIg

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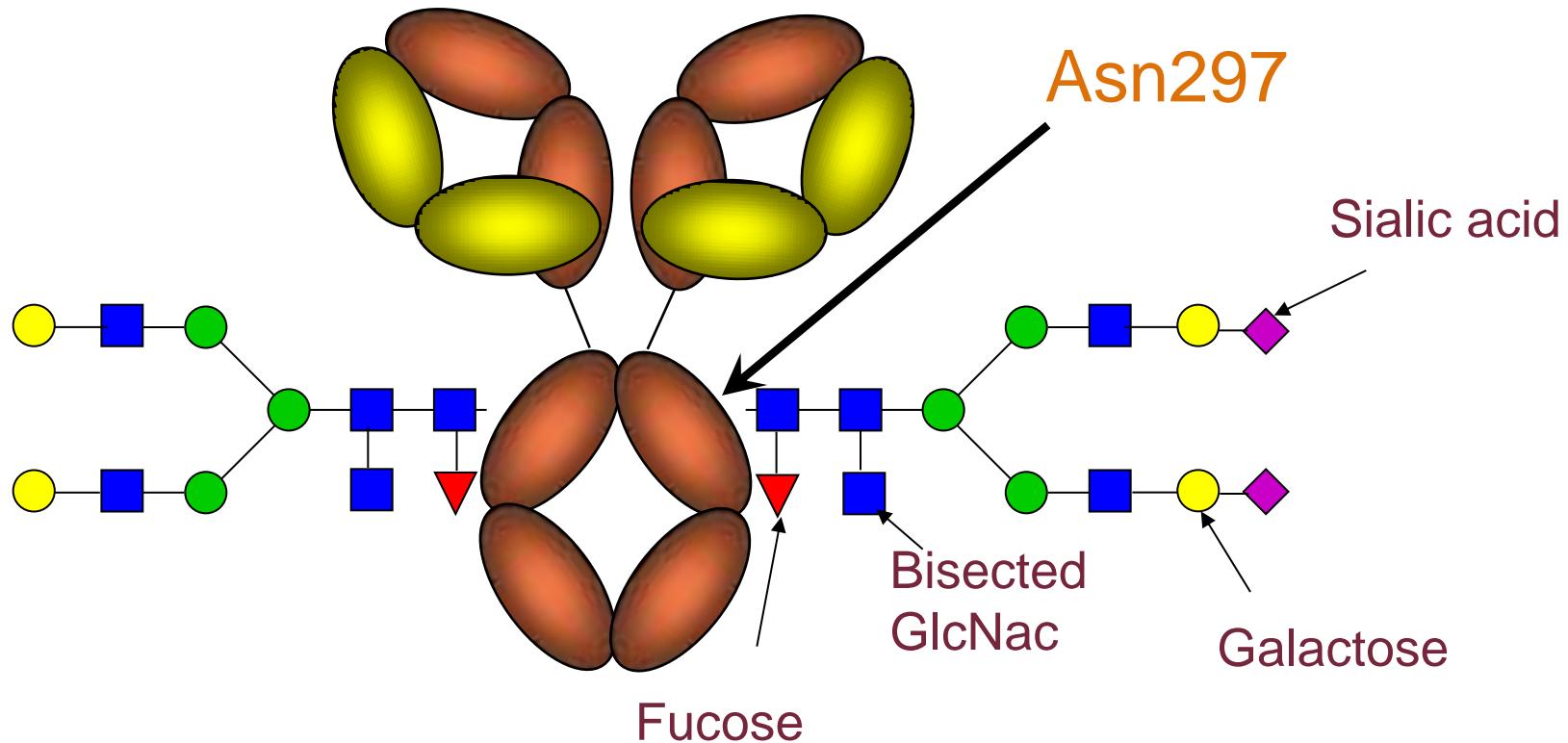
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Increasing the efficiency of RhIg

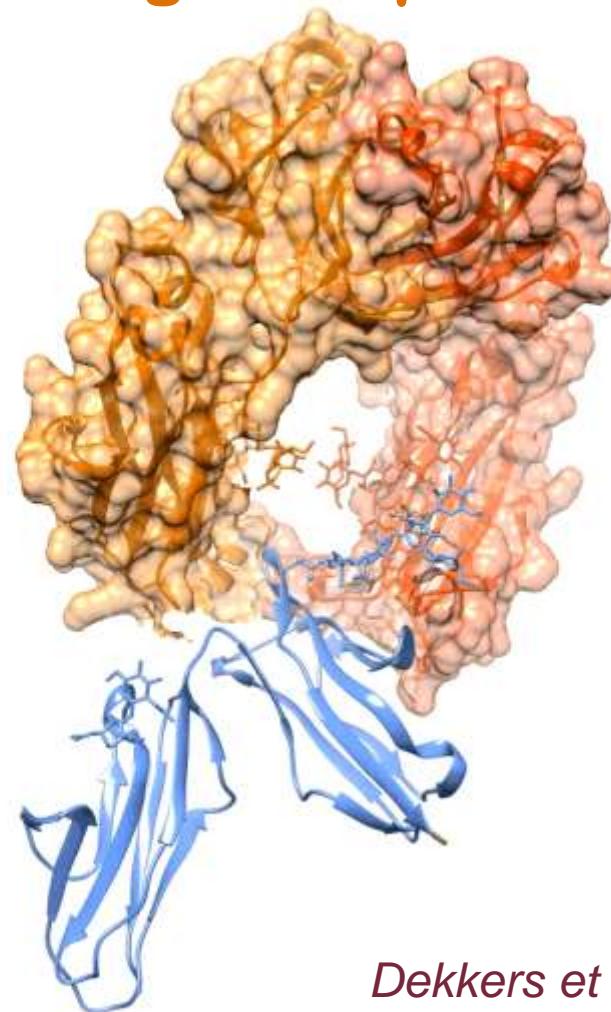
1. Increasing the biological activity of RhIg
 - Effect of Fc-core glycosylation of anti-D immunoglobulin
2. Decreasing the unnecessary use of RhIg
 - Guiding of immunoprophylaxis by fetal RHD typing

Fc-glycans



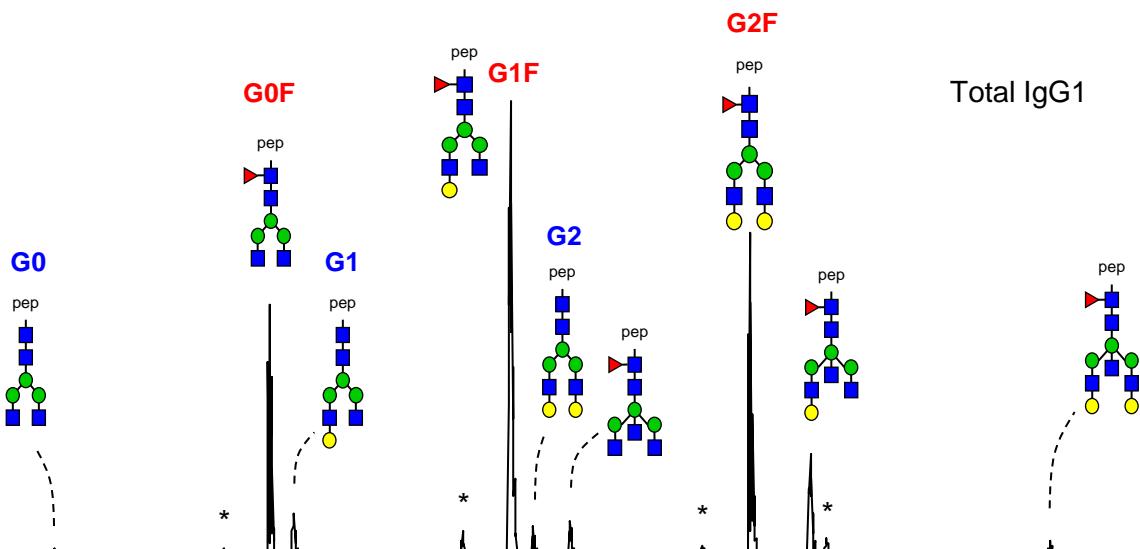
Glycans affect binding to Fc γ R

Low Fc-fucosylation
results in **50x**
enhanced binding
To Fc γ RIIIa



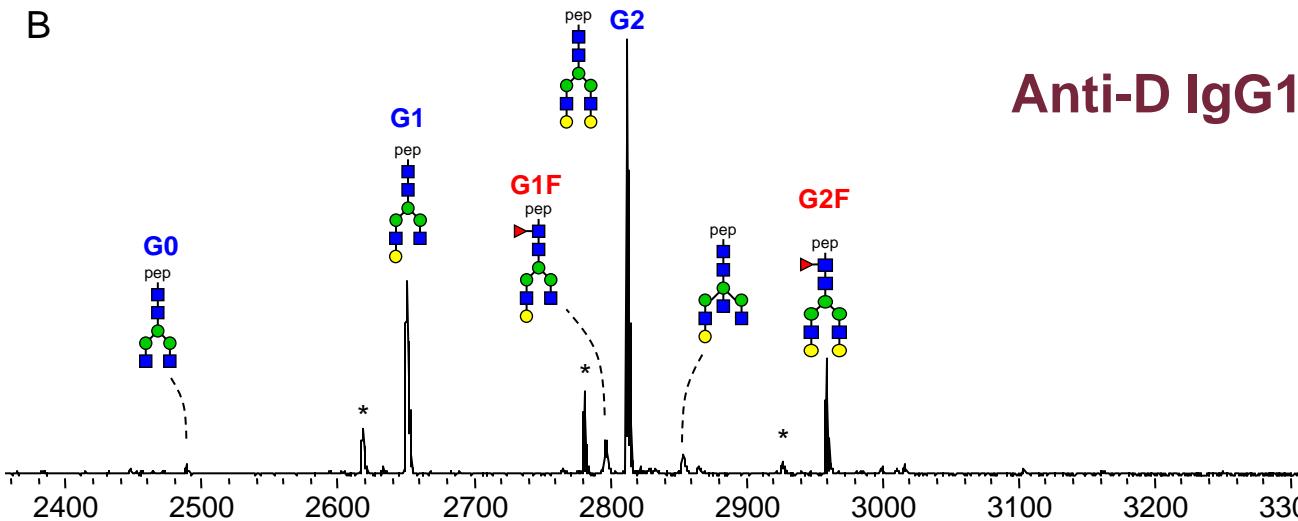
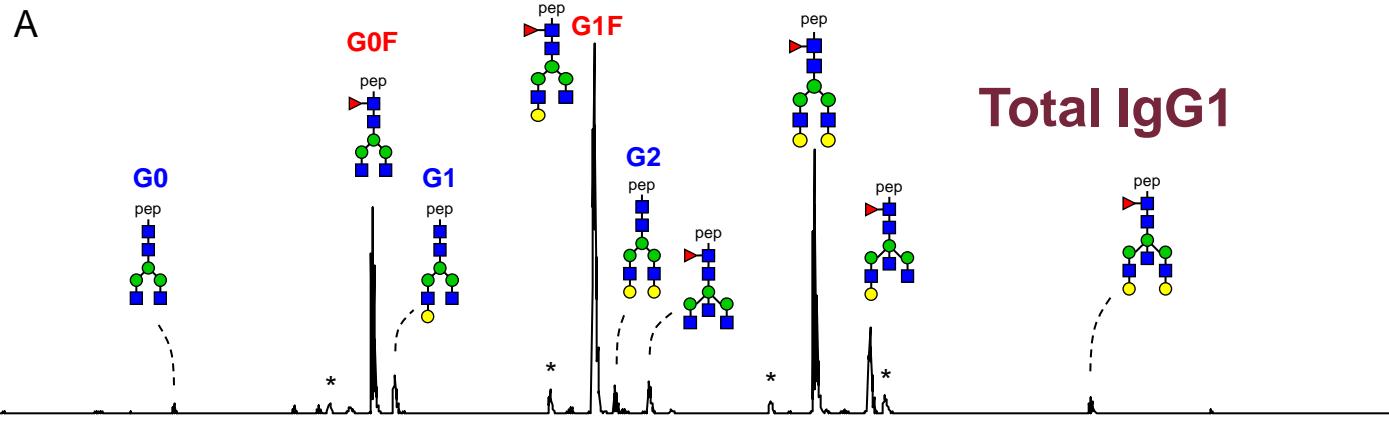
Fc glycopeptides: Mass spectrometric analysis

Glycan species	Glycan structure	IgG1 E ₂₉₃ EQYNSTYR ₃₀₁ ^a P01857 ^b [M+H] ⁺
No glycan	-	1190.5198
G0		2487.9880
G0F		2634.0459
G1		2 α A
G1F		2796.0987
G0N		2691.0674
G0FN		2837.1253
G2		2812.0936
G2F		2958.1515
G1N		2853.1202
G1FN		2999.1781
G2N		3015.1730
G2FN		3161.2309

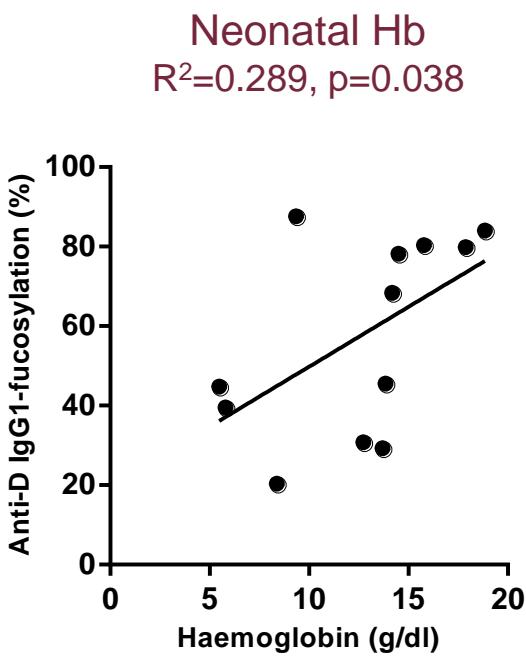
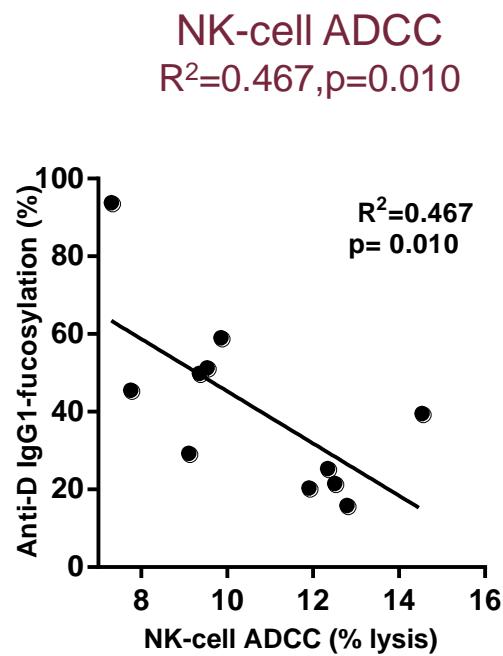
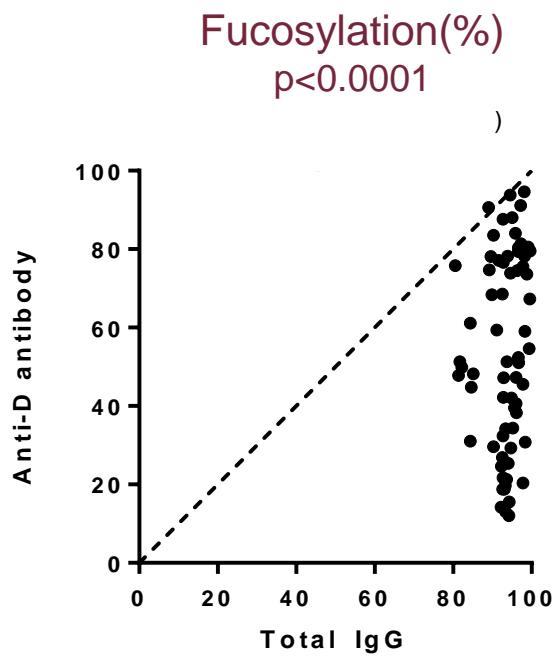


Majority of IgG1 in plasma is fucosylated

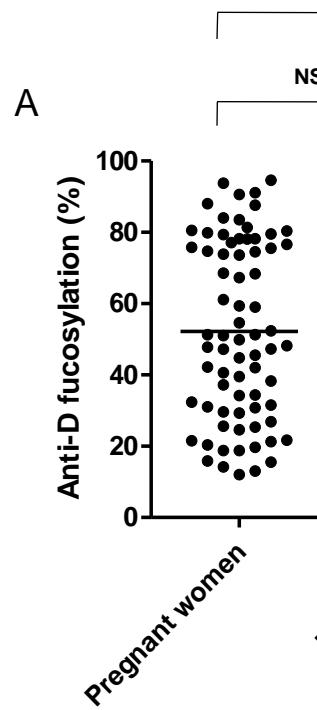
Anti-D IgG is low - fucosylated



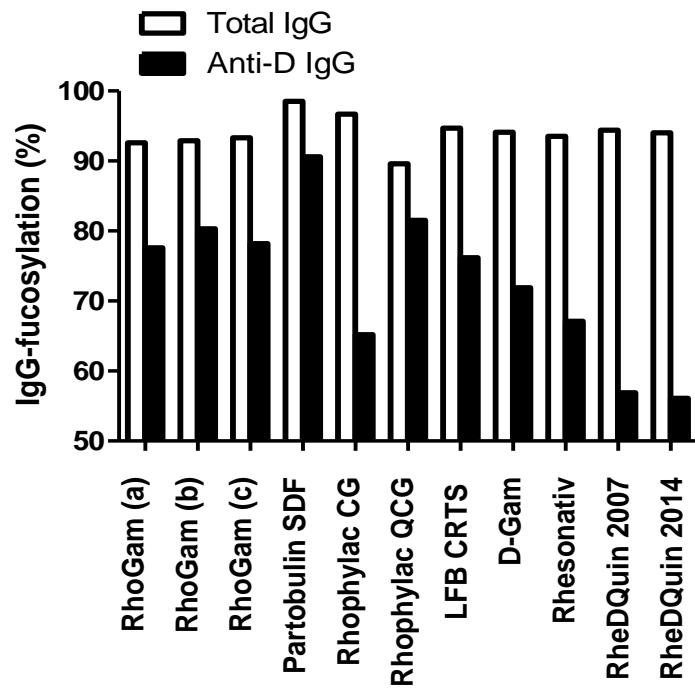
Fc-fucosylation of anti-D IgG is variable and related to pathogenicity



Fc-fucosylation of anti-D IgG from hyperimmune donors is variably low



Anti-D preparations display variable decreases in Fc-fucosylation



Efficacy of anti-D appears to be related to Fc-fucosylation

- *Human polyclonal anti-D immunoglobulin:*
 - **Low** level of fucosylation:
 - high ADCC, very rapid RBC clearance, **prevented** D-immunization.
- *Moab anti-D produced in human B cell lines (BRAD3, BRAD5)*
 - **Medium** level of fucosylation
 - medium ADCC, fast clearance, prevented D-immunization **in 93%** subjects
 -
- *Moab anti-D produced in human-mouse hetero hybridoma: (Fog1, AD1, G7, G12)*
 - **High** level of fucosylation
 - low ADCC, variable and slow clearance, **stimulated** D-immunization.

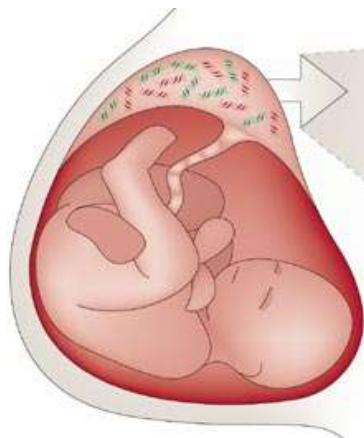
Conclusion (1)

Production of RhIg from hyperimmune donors with low fucosylated anti-D Ig might result in more potent RhIg

Guiding Rh immunoprophylaxis by fetal *RHD* typing

- 40% of D-neg women carry a D-neg child and do not need Rh-Ig
- Fetal DNA is present in low concentration in maternal plasma from week 5

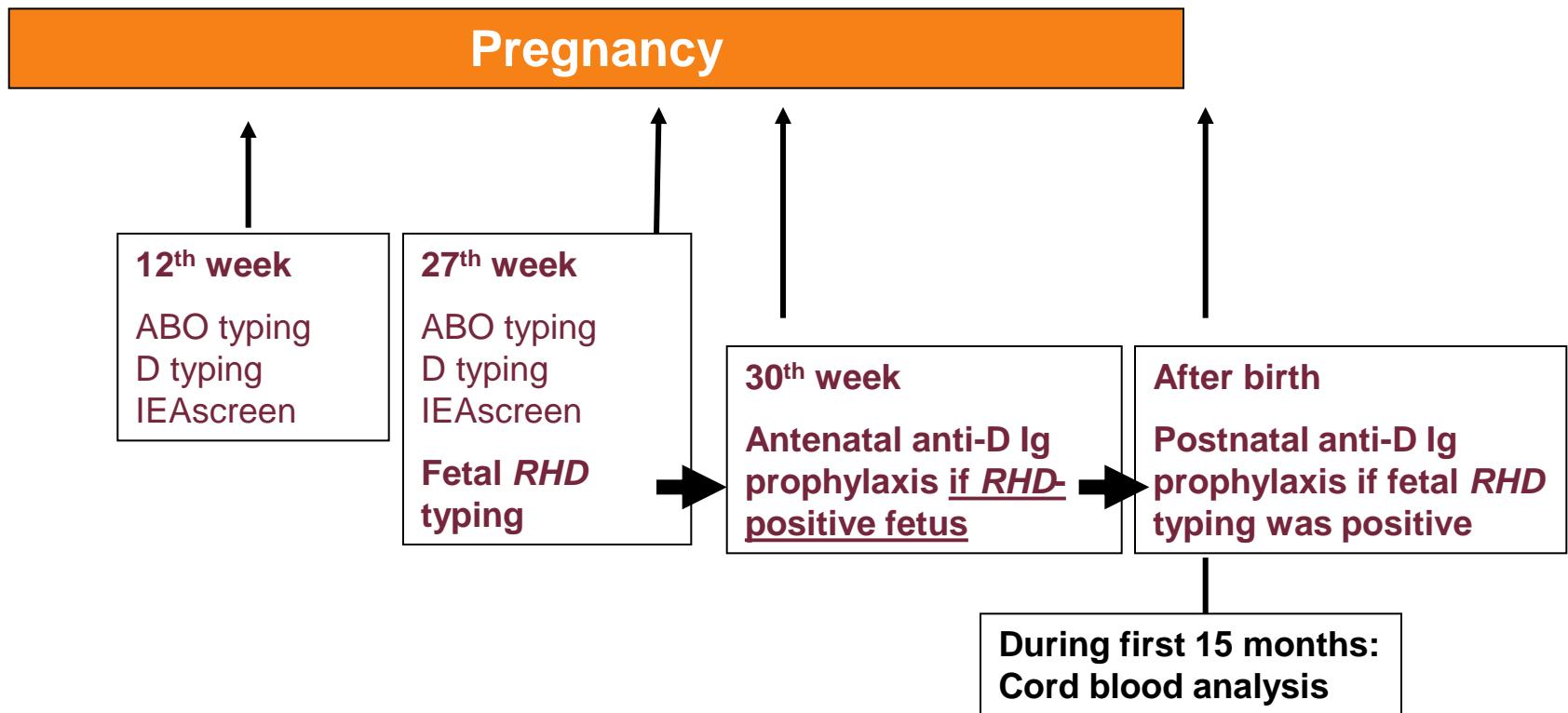
placenta → plasma
of pregnant woman



Mixture of DNA
from **mother** and
DNA from **fetus**

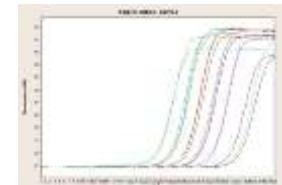
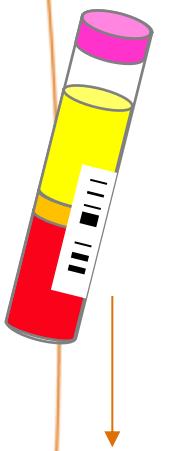
Illustration: Lo, 2007

Dutch screening program



Technical approach

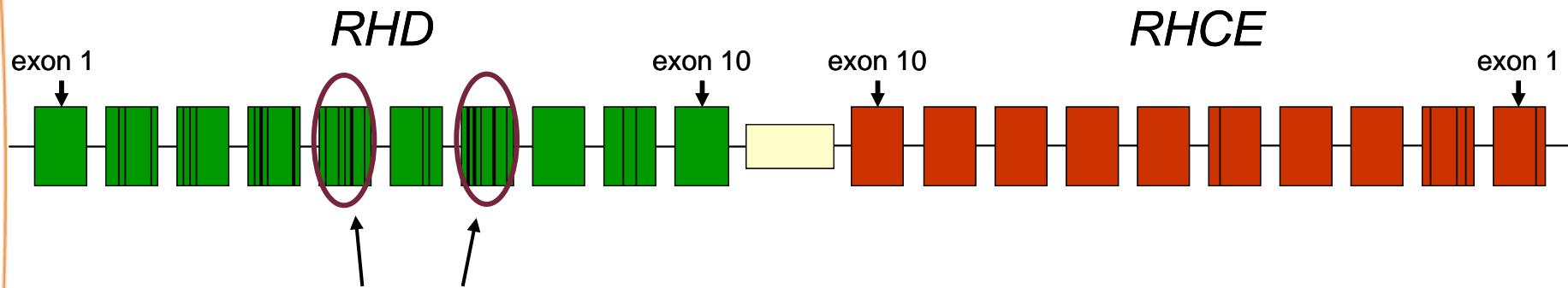
- Centralized at one laboratory (Sanquin, Amsterdam)
- DNA isolation from 1 ml of EDTA plasma in **27th week** of pregnancy
- First 15 months comparison with cord blood



Electronic
Result

Report

Design fetal *RHD* typing



RHD-PCR Multiplex

exon 5: not amplified in majority of *RHD* variants

in Caucasians: RHD*DVI

in Blacks: RHD*Ψ; RHD*01N.06 and RHD*03N.01 (r's)

exon 7: present in most *RHD* variants



First 25.500 samples: Diagnostic accuracy of at least 99.1%

CB-serology	Pos	Neg
Plasma-PCR		
Pos	15,826	225 (0.87%)
Neg	9 (0.03%)	9,740

Systemic analysis of false results

- **False negative fetal RHD typing:**

- No antenatal prophylaxis : 0.61% risk of alloimmunization
- No postnatal prophylaxis: 17% risk of alloimmunization

- **False positive fetal RHD typing:**

- Unnecessary RhIg is given, no clinical consequences

Potential effect of PCR guided ante- and postnatal immunoprophylaxis

	Unnecessary antenatal anti-D	No antenatal anti-D, while at risk	No postnatal anti-D, while at risk
Old program (no PCR, only cord blood)	38,3%	0%	0,09%
New program (only PCR, no cord blood)	0,43%	0,03%	0,03%

Accuracy of implemented fetal RHD typing (Denmark, the Netherlands, Finland)

	Samples	TRUE Pos	False Pos	TRUE Neg	FALSE Neg	sensitivity %	specificity %
	Clausen et al., 2014	12688	7636	41	4706	11	99.86
	de Haas et al., 2016	25789	15816	225	9739	9	99.94
	Haimila et al., 2017	10814	7080	7	3640	1	99.99
Total		49291			21	99.93	

van der Schoot et al. Curr Opin Hem 2017

Conclusion (2)

Implementation of fetal *RHD* typing has resulted in decreased unnecessary use of antenatal RhIg, without loss of efficiency of the program

Acknowledgements

- Fc-glycosylation
 - **Gestur Vidarsson**
 - Rick Kapur
 - Gillian Dekkers
 - Lussy Della Valle
 - Myrthe Sonneveld
 - **Manfred Wührer** (LUMC, Leiden)
 - Belinda Kumpel (Bristol)
- Fetal RHD genotyping
 - **Masja de Haas**
 - **Barbera Veldhuisen**
 - Florentine Thurik
 - Peter Scheffer
 - Aicha Ait Soussan
 - Tamara Stegman
 - Lieve Christiaens (UMCU, Utrecht)